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PATENT COOPERATION TREATY

From the INTERNATIONAL BUREAU

PCT

NOTIFICATION OF ELECTION

(PCT Rule 61.2)

To:

United States Patent and Trademark Office
(Box PCT)
Crystal Plaza 2
Washington, DC 20231
ETATS-UNIS D'AMERIQUE

in its capacity as elected Office

Date of mailing (day/month/year) 30 September 1996 (30.09.96)	
International application No. PCT/AU96/00149	**Applicant's or agent's file reference** IRN444950
International filing date (day/month/year) 15 March 1996 (15.03.96)	**Priority date** (day/month/year) 16 March 1995 (16.03.95)
Applicant WALKER, John et al	

1. The designated Office is hereby notified of its election made:

   [X] in the demand filed with the International Preliminary Examining Authority on:

   04 September 1996 (04.09.96)

   [ ] in a notice effecting later election filed with the International Bureau on:

   _____

2. The election [X] was

   [ ] was not

   made before the expiration of 19 months from the priority date or, where Rule 32 applies, within the time limit under Rule 32.2(b).

The International Bureau of WIPO
34, chemin des Colombettes
1211 Geneva 20, Switzerland

Facsimile No.: (41-22) 740.14.35

Authorized officer:
Marie-José Devillard

Telephone No.: (41-22) 730.91.11

PATENT COOPERATION TREATY

From the
INTERNATIONAL PRELIMINARY EXAMINING AUTHORITY

To: Agent :
PHILLIPS ORMONDE & FITZPATRICK
367 Collins Street
MELBOURNE VIC 3000

PCT PUT. SDI/F

NOTIFICATION OF RECEIPT OF DEMAND

(PCT Rule 61.1(b), first sentence and Administrative Instructions, Section 601)

Date of mailing *(day/month/year)*	19 SEP 1996 (19/9/96)
Applicant's or agent's file reference: 444950	IMPORTANT NOTIFICATION

International application No.	International filing date *(day/month/year)*	Priority date *(day/month/year)*
PCT/AU96/00149	15 MAR 1996 (15/3/96)	16 MAR 1995 (16/3/95)

Applicant: University of Melbourne; The (et al.)

1. The applicant is hereby notified that this International Preliminary Examining Authority considers the following date as the date of receipt of the demand for international preliminary examination of the international application:

4 SEP 1996 (4/9/96)

RECEIVED 18 SEP 1996
PHILLIPS ORMONDE & FITZPATRICK
SIGNED
FOR COMPUTER ENTRY
COMPUTER MOS
REFERRED TO MOS

2. This date of receipt is:

- [X] the actual date of receipt of the demand.
- [ ] the date on which the proper corrections to the demand were timely received.

3. [ ] This date is AFTER the expiration of 19 months from the priority date.

Attention: The election(s) made in the demand does (do) not have the effect of postponing the commencement of the national phase until 30 months from the priority date (or later in some Offices) (Article 39(1)). Therefore, the acts for entry into the national phase must be performed within 20 months from the priority date (or later in some Offices) (Article 22).

[ ] This notification confirms the information given in person or by telephone on: _____

4. Only where paragraph 3 applies, a copy of this notification has been sent to the International Bureau.

Name and mailing address of the IPEA/AU AUSTRALIAN INDUSTRIAL PROPERTY ORGANISATION P.O. BOX 200, WODEN, A.C.T. 2606 AUSTRALIA Facsimile No. 06 285 3929	Authorized officer: L. Tracey Telephone No. 2832511

Form PCT/IPEA/402 (July 1992)

INTERNATIONAL SEARCH REPORT

International Application No.
PCT/AU 96/00149

A. CLASSIFICATION OF SUBJECT MATTER

Int Cl[6]: C07K 16/12; C12N 15/31; C12P 21/00, 21/02, 21/08; G01N 33/53, 33/531; A61K 39/04; C07K 14/30

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)
IPC As Above

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched
AU: IPC As Above

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)
WPAT, BIOT, JAPIO: MYCOPLASM: AND ANTIGEN#
CASM: MULTI-SEQUENCES

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	AU,A, 70685/87 (CETUS CORP) 1 October 1987 Example 1, Claims	1,16,17,20-23,33 35-37,43
X	AU,B, 49035/90 (640364) (UNIVERSITY OF MELBOURNE et al) 11 October 1990 Claims	1,2,16,20,22
X	AU,A, 76820/91 (SYNERGEN, INC.) 17 October 1991 Fig. 1, 4, 6 and 7, Claim 3	14,38,39,41,42

[X] Further documents are listed in the continuation of Box C

[X] See patent family annex

* Special categories of cited documents:

"A" document defining the general state of the art which is not considered to be of particular relevance

"E" earlier document but published on or after the international filing date

"L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)

"O" document referring to an oral disclosure, use, exhibition or other means

"P" document published prior to the international filing date but later than the priority date claimed

"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention

"X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone

"Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art

"&" document member of the same patent family

Date of the actual completion of the international search	Date of mailing of the international search report
6 May 1996	15.05.96

Name and mailing address of the ISA/AU	Authorized officer
AUSTRALIAN INDUSTRIAL PROPERTY ORGANISATION PO BOX 200 WODEN ACT 2606 AUSTRALIA Facsimile No.: (06) 285 3929	BARRY SPENCER Telephone No.: (06) 283 2234

INTERNATIONAL SEARCH REPORT

International Application No.
PCT/AU 96/00149

C (Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
PX	AU,A, 17602/95 (SYNERGEN, INC.) 26 October 1995 Fig. 1, 4, 6 and 7 P 13 L14-P14 L5	14,38,39,41,4
X	AU,B, 49599/90 (633970) (AUSPHARM INTERNATIONAL LTD.) 26 July 1990 Examples 4 and 5	16,17,20,21,3
X	US,A, 4894332 (SCHALLER et al) 16 January 1990 Examples	1,2,14,16,17, 20-22,24,33, 35,37-39,41,4
X	US,A, 5252328 (FAULDS et al) 12 October 1993 Examples 1 to 4	1-17,20-25, 33-35,37
X	US,A, 5240706 (FAULDS) 31 August 1993 Examples 1 and 2, Claims	1-6,14-16,21 22,24,25,33-3 37,38,41,42
X	EP,A, 0475185 (NIPPON FLOUR MILLS CO., LTD.) Refer Example 1, Claims	1-6,12-17, 20-25,33-35, 37-39,41,42
PX	Journal of Bacteriology, Vol. 177, No. 7, April 1995, pp1915-1917, "Molecular Cloning of a 46-Kilodalton Surface Antigen (P46) Gene from Mycoplasma hyopneumoniae: Direct evidence of CGG Codon Usage for Arginine", Futo et al	3-6,12-15,24 25,31,32,38, 39,41,42
PX	Journal of Clinical Microbiology, Vol. 33, No. 3, March 1995, pp 680-683, "Recombinant 46-Kilodalton Surface Antigen (P46) of Mycoplasma hyopneumoniae Expressed in Escherichia Coli Can Be Used for early Specific Diagnosis of Mycoplasmal Pneumonia of Swine by Enzyme-Linked Immunosorbent assay", Futo et al	3-6, 12-15,38,39, 41,42
X	Infection and Immunity, Vol. 49, No. 2, pp329-335, "Surface Proteins of Mycoplasma hyopneumoniae Identified from an Escherichia coli Expression Plasmid Library", Klinkert et al	3-6,12-15,38 39,41,42
A	EP,A, 0571648 (WENG) 1 December 1993	
A	Derwent abstract Accession No. 88-010509, Class S03, JP,A, 62-275455 (NORINSHO KK) 27 November 1987	
A	Derwent Abstract Accession No. 90-241949, Class S03, JP,A, 02-167079 (NIPPON SEIFUN KK) 27 June 1990	
A	Derwent abstract Accession No. 95-203749, Class B04, C06, D16, JP,A, 07-118167 (ZENKOKU NOGYO KYODO KUMIAI RENGOKAI) 9 May 1995	

INTERNATIONAL SEARCH REPORT

This Annex lists the known "A" publication level patent family members relating to the patent documents cit in the above-mentioned international search report. The Australian Patent Office is in no way liable for thes particulars which are merely given for the purpose of information.

Patent Document Cited in Search Report		Patent Family Member					
AU	87/70685	EP	254384	JP	63000298		
AU	90/49035	AT	134386	BR	9000451	CA	2008808
		CN	1046188	DE	69025414	EP	381427
		FI	900498	IL	93234	JP	3087199
		NO	178893	NZ	232279	ZA	9000766
AU	91/76820	AU	17602/95	CA	2078131	EP	527771
		FI	924428	HU	65827	JP	5506984
		NO	923828	WO	9115593	US	5459048
		AT	135048	AU	622855	DE	3751727
		DK	1608/88	EP	315637	HU	208550
		IL	83324	JP	1503735	WO	8800977
AU	95/17602	AU	76820/91	CA	2078131	EP	527771
		FI	924428	HU	65827	JP	5506984
		NO	923828	WO	9115593	US	5459048
		AT	135048	AU	622855	DE	3751727
		DK	1608/88	EP	315637	HU	208550
		IL	83324	JP	1503735	WO	8800977
AU	90/49599	EP	454735	NZ	232190	WO	9007935
		ZA	9000474				
US	4894332	CA	1301677	CN	86102858	EP	196215
		JP	61274687				

END OF AN

INTERNATIONAL SEARCH REPORT

International Application No
PCT/AU 96/00149

This Annex lists the known "A" publication level patent family members relating to the patent documents cit in the above-mentioned international search report. The Australian Patent Office is in no way liable for thes particulars which are merely given for the purpose of information.

Patent Document Cited in Search Report		Patent Family Member					
US	5252328	CA	1321142	CN	88101554	DK	1674/88
		EP	283840	HU	203672	IE	61626
		JP	63258427	PT	87041		
US	5240706	AT	134705	DE	68925769	EP	359919
		JP	2291271				
EP	475185	JP	5091882				

END OF ANN

PATENT COOPERATION TREATY

PCT

INTERNATIONAL PRELIMINARY EXAMINATION REPORT

(PCT Article 36 and Rule 70)

Applicant's or agent's file reference IRN 444950	FOR FURTHER ACTION	See Notification of Transmittal of International Preliminary Examination Report (Form PCT/IPEA/416).
International application No. PCT/AU 96/00149	International filing date 15 March 1996	Priority Date 16 March 1995

International Patent Classification (IPC) or national classification and IPC

Int. Cl.[6] C07K 16/12, 14/30; C12N 15/31; C12P 21/00, 21/02, 21/08; G01N 33/53, 33/531; A61K 39/04

Applicant
(1) UNIVERSITY OF MELBOURNE
(2) WALKER, John; LEE, Rogan; DOUGHTY, Stephen William

1. This international preliminary examination report has been prepared by this International Preliminary Examining Authority and is transmitted to the applicant according to Article 36.

2. This REPORT consists of a total of 6 sheets, including this cover sheet.

☐ This report is also accompanied by ANNEXES, i.e., sheets of the description, claims and/or drawings which have been amended and are the basis for this report and/or sheets containing rectifications made before this Authority (see Rule 70.16 and Section 607 of the Administrative Instructions under the PCT).

These annexes consist of a total of sheet(s).

3. This report contains indications relating to the following items:

I	☒	Basis of the report
II	☐	Priority
III	☐	Non-establishment of opinion with regard to novelty, inventive step and industrial applicability
IV	☒	Lack of unity of invention
V	☒	Reasoned statement under Article 35(2) with regard to novelty, inventive step or industrial applicability; citations and explanations supporting such statement
VI	☒	Certain documents cited
VII	☐	Certain defects in the international application
VIII	☒	Certain observations on the international application

Date of submission of the demand	Date of completion of the report
4 September 1996	23 December 1996
Name and mailing address of the IPEA/AU AUSTRALIAN INDUSTRIAL PROPERTY ORGANISATION PO BOX 200 WODEN ACT 2606 AUSTRALIA Facsimile No. (06) 285 3929	Authorized Officer BARRY SPENCER Telephone No. (06) 283 2284

I Basis of the report

1. This report has been drawn on the basis of *(Replacement sheets which have been furnished to the receiving Office in response to an invitation under Article 14 are referred to in this report as "originally filed" and are not annexed to the report since they do not contain amendments.)*:

- [X] the international application as originally filed.
- [ ] the description, pages , as originally filed,
  pages , filed with the demand,
  pages , filed with the letter of ,
  pages , filed with the letter of .
- [ ] the claims, Nos. , as originally filed,
  Nos. , as amended under Article 19,
  Nos. , filed with the demand,
  Nos. , filed with the letter of ,
  Nos. , filed with the letter of .
- [ ] the drawings, sheets/fig , as originally filed,
  sheets/fig , filed with the demand,
  sheets/fig , filed with the letter of ,
  sheets/fig , filed with the letter of .

2. The amendments have resulted in the cancellation of:

- [ ] the description, pages
- [ ] the claims, Nos.
- [ ] the drawings, sheets/fig

3. [ ] This report has been established as if (some of) the amendments had not been made, since they have been considered to go beyond the disclosure as filed, as indicated in the Supplemental Box (Rule 70.2(c)).

4. Additional observations, if necessary:

IV. Lack of unity of invention

1. In response to the invitation to restrict or pay additional fees the applicant has:

- [ ] restricted the claims.
- [ ] paid additional fees.
- [ ] paid additional fees under protest.
- [ ] neither restricted nor paid additional fees.

2. [X] This Authority found that the requirement of unity of invention is not complied with and chose, according to Rule 68.1, not to invite the applicant to restrict or pay additional fees.

3. This Authority considers that the requirement of unity of invention in accordance with Rules 13.1, 13.2 and 13.3 is

- [ ] complied with.
- [X] not complied with for the following reasons:

Claims 41 and 42 are not so limited to an amino acid sequence of a Mycoplasma as to constitute the same invention as claimed in claims 1-40 and 43.

4. Consequently, the following parts of the international application were the subject of international preliminary examination in establishing this report:

- [X] all parts.
- [ ] the parts relating to claims Nos.

INTERNATIONAL PRELIMINARY EXAMINATION REPORT

International application No.
PCT/AU 96/00149

V. Reasoned statement under Article 35(2) with regard to novelty, inventive step or industrial applicability; citations and explanations supporting such statement

1. Statement

Novelty (N)	Claims	1-43	YES
	Claims		NO
Inventive step (IS)	Claims	7-11, 18, 19, 25-32, 34, 40	YES
	Claims	1-6, 12-17, 20-24, 33, 35-39, 41-43	NO
Industrial applicability (IA)	Claims	1-43	YES
	Claims		NO

2. Citations and explanations

D1 AU,A, 70685/87

D2 AU,B, 49035/90 (640364)

D3 AU,A, 76820/91

D4 AU,A, 17602/95

D5 AU,B, 49599/90 (638970)

D6 US,A, 4894332

D7 US,A, 5252328

D8 US,A, 5240706

D9 EP,A, 0475185

D10 Journal of Bacteriology, Volume 177, No. 7, pages 1915-1917

D11 Journal of Clinical Microbiology, Volume 33, No. 3, pages 680-683

D12 Infection and Immunity, Volume 49, No. 2, pages 329-335

Claims 1-6, 12-17, 20-24, 33, 35-39, 41-43 do not involve an inventive step when compared to D2, when read in the light of D1, D3-D12. D1, D3-D12 show that before the priority date, the use of antigens targeted to surface lipoproteins or membrane proteins of Mycoplasma were known and sought after. D2 discloses the exact same method for a group of pathogens, one of which can be mycoplasmas. A PSA having regard to all these documents would find no burden of experimentation or inventive faculty to use the same method as D2 for obtaining antigens to mycoplasmas

D1 discloses taking a biological sample three days after the last injection which would be included within your term "sho. time".

D10-D12 all disclose the nucleic acid sequences as set out in claims 14, 15, 38 and 42.

VI. Certain documents cited

1. Certain published documents (Rule 70.10)

Application No. Patent No.	Publication date *(day/month/year)*	Filing date *(day/month/year)*	Priority date (valid claim *(day/month/year)*
AU,A, 17602/95	26 October 1995	21 April 1995	2 April 1990

2. Non-written disclosures (Rule 70.9)

Kind of non-written disclosure	Date of non-written disclosure *(day/month/year)*	Date of written disclosure referring to non-written disclosure *(day/month/year)*

VIII Certain observations on the international application

The following observations on the clarity of the claims, description, and drawings or on the question whether the claims are fully supported by the description, are made:

Claims 1 and 16 lack clarity in their reference to "short time". It is unclear what time frame is included by this term and the description gives no further indication as to any limits.

Claims 41 and 42 are not fully supported by the description because the description is directed towards Mycoplasma yet these claims are to any amino acid sequence.

PCT WORLD INTELLECTUAL PROPERTY ORGANIZATION International Bureau

U 011415-0

INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(51) International Patent Classification 6: C07K 16/12, C12N 15/31, C12P 21/00, 21/02, 21/08, G01N 33/53, 33/531, A61K 39/04, C07K 14/30 — A1

(11) International Publication Number: WO 96/28472

(43) International Publication Date: 19 September 1996 (19.09.96)

(21) International Application Number: PCT/AU96/00149

(22) International Filing Date: 15 March 1996 (15.03.96)

(30) Priority Data:
PN 1789 16 March 1995 (16.03.95) AU

(71) Applicant (*for all designated States except US*): THE UNIVERSITY OF MELBOURNE [AU/AU]; Grattan Street, Parkville, VIC 3052 (AU).

(72) Inventors; and

(75) Inventors/Applicants (*for US only*): WALKER, John [AU/AU]; 26 Clapham Street, Balwyn, VIC 3103 (AU). LEE, Rogan [AU/AU]; 73 Greenford Street, Chapel Hill, QLD 4069 (AU). DOUGHTY, Stephen, William [AU/AU]; 1A Diana Drive, Blackburn, VIC 3103 (AU).

(74) Agent: PHILLIPS ORMONDE & FITZPATRICK; 367 Collins Street, Melbourne, VIC 3000 (AU).

(81) Designated States: AL, AM, AT, AU, AZ, BB, BG, BR, BY, CA, CH, CN, CZ, DE, DK, EE, ES, FI, GB, GE, HU, IS, JP, KE, KG, KP, KR, KZ, LK, LR, LS, LT, LU, LV, MD, MG, MK, MN, MW, MX, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, TJ, TM, TR, TT, UA, UG, US, UZ, VN, ARIPO patent (KE, LS, MW, SD, SZ, UG), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, CH, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, ML, MR, NE, SN, TD, TG).

Published
With international search report.

(54) Title: ANTIGEN COMPOSITION AGAINST MYCOPLASMA

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          10         20         30         40         50
1234567890 1234567890 1234567890 1234567890 1234567890

MKKMLRKKFL YSSAIYATSL ASIIAFVAAG CGQTESGSTS DSKPQAETLK      50
HKVSNDSIRI ALTDPDNPRW ISAQKDIISY VDETEAATST IIKNQDAQNN     100
WLTQQANLSP APKGFIIAPE NGSGVGTAVN TIADKGIPIV AYDRLITGSD     150
KYDWYVSFDN EKVGELQGLS LAAGLLGKED GAFDSIDQMN EYLKSHMPQE     200
TISFYTIAGS QDDNNSQVFY NGAMKVLKEL MKNSQNKIID LSPEGENAVY     250
VPGWNYGTAG QRIQSFLTIN KDPAGGNKIK AVGSKPASIF KGFLAPNDGM     300
AEQAITKLKL EGFDIQKIFV TRQDYNDKAK TFIKDGDQNM TIYKPDKVLG     350
KVAVEVLRVL IAKQNKASRS EVENEIKAKL PNISFKYDNQ TYKVQGKNIN     400
TILVSPVIVT KANVDNPDA                                       419
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(57) Abstract

The present invention relates to a putative protective antigen against a *Mycoplasma*, prepared by a method including a sample of a *Mycoplasma*; an antibody probe including at least one antibody against a *Mycoplasma* produced by a method including: providing a biological sample taken a short time after an immune animal has been challenged with a *Mycoplasma* or *Mycoplasma* extract taken from the infection site or an area of a lesion or an area close to the infection site or lesion; isolating cells from the biological sample; culturing cells *in vitro* in a suitable culture medium; and harvesting antibodies produced from said cells; probing the *Mycoplasma* sample with the antibody probe to detect at least one antigen; and isolating the antigen detected, also including diagnostic antigens, the preparation thereof, and their use in the formation of vaccine compositions, particularly vaccine compositions against *Mycoplasma hyopneumoniae* infections.

52 Mycoplasmas

Shmuel Razin, PhD

General Concepts

Mycoplasmas are the smallest and simplest self-replicating bacteria. The mycoplasma cell contains only the minimum set of organelles essential for growth and replication: a plasma membrane, ribosomes, and a double-stranded DNA molecule (Figure 52-1). The relative simplicity of mycoplasmas has made them most useful tools for studying basic problems in cell biology. Unlike all other prokaryotes, the mycoplasmas have no cell walls and are placed accordingly in a separate class: Mollicutes (*mollis*, soft; *cutis*, skin). The common term mycoplasmas is used rather loosely to denote any species included in the class Mollicutes, whereas the common names acholeplasmas, ureaplasmas, spiroplasmas, and anaeroplasmas are used to refer to members of the corresponding genera, rather than to defined species within the genera (Table 52-1).

All mycoplasmas cultivated and identified so far are parasites of humans, animals, plants, and arthropods. Primary habitats of animal mycoplasmas are the mucous surfaces of respiratory and urogenital tracts and the joints in some animals. Although some

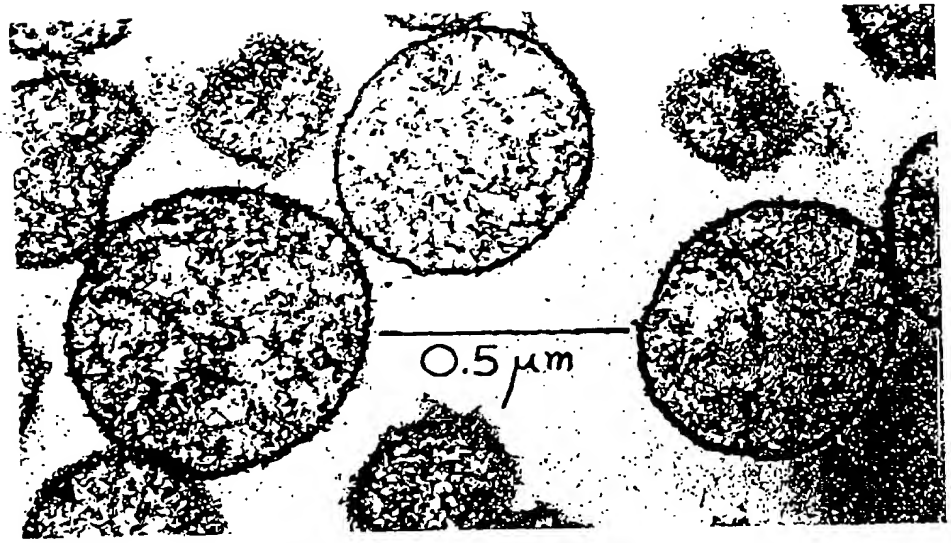

Figure 52-1 Electron micrograph of thin sectioned mycoplasma cells. Cells are bounded by a single membrane showing in section the characteristic trilaminar shape. Cytoplasm contains thin threads representing sectioned chromosome and dark granules representing ribosomes. Courtesy R.M. Cole

Table 52-1 Taxonomy and Properties of Organisms Included in the Class Mollicutes

Classification	*Number of Recognized Species*	*Genome*		*Cholesterol Requirement*	*Characteristic Properties*	*Habitat*
		Size ($\times 10^6$ daltons)	*G+C Content (%)*			
Mycoplasmataceae						
Mycoplasma	about 70	5	23–41	+	—	Animals and humans
Ureaplasma	2	5	28	+	Urease activity	Animals and humans
Acholeplasmataceae						
Acholeplasma	9	10	27–35	−	—	Animals and humans
Spiroplasmataceae						
Spiroplasma	4	10	26–30	+	Helical filaments	Arthropods and plants
Genus of uncertain taxonomic position						
Anaeroplasma	2	ND	29–34	Some + some −	Anaerobic; some digest bacteria	Rumens of cattle and sheep

mycoplasmas belong to the normal flora, most species are pathogens, causing various diseases that tend to run a chronic course. In humans, one species, *Mycoplasma pneumoniae*, has been established as the agent of primary atypical pneumonia; another species, *Ureaplasma urealyticum*, probably is associated with nongonococcal urethritis. Many mycoplasma species are established pathogens of farm animals, causing contagious pleuropneumonia and mastitis in cattle and goats and chronic respiratory diseases and arthritis in swine, chicken, and laboratory animals. In plants, mycoplasmas inhabit the phloem tissue and cause a large variety of economically important diseases in crops.

The pathogenic mechanisms of mycoplasmal infections are not clearly understood, but adherence to host cells by protein adhesins appears to be involved. The role of toxic factors (H_2O_2, NH_3) in the disease process is under investigation. Infection with *M pneumoniae* occurs worldwide and is transmitted by respiratory droplets, whereas *U urealyticum* is transmitted by sexual contact. Mycoplasmal infections can be treated effectively with broad-spectrum antibiotics such as tetracycline.

Mycoplasmas have been nicknamed the crabgrass of cell cultures because their infections are persistent, difficult to cure, and frequently difficult to detect and diagnose. Contamination by mycoplasmas presents serious problems in the production of viral vaccines in cell cultures. The origin of contaminating mycoplasmas is in components of the cell culture medium, particularly serum, or from the mycoplasma flora of the technician's mouth, spread by droplet infection.

Distinctive Properties

Morphology and Reproduction

The coccus is the basic form of all mycoplasma cultures. The diameter of the smallest cocci capable of reproduction is about 300 nm. Hence, many of the minute and plastic cells can be squeezed through membrane filters of 450 nm pore diameter. In most mycoplasma cultures, elongated or filamentous forms (up to 100 μm long and about 0.4 μm thick) also can be observed (Figure 52-2). The filaments tend to produce truly

Figure 52-2 Scanning electron micrograph of a *M pneumoniae* culture, showing characteristic morphologic elements, including chains of cocci, branched filaments, and elongated cells with tapered tips (specialized tip structures). From Biberfeld, G, Biberfeld, P.: *J Bacteriol* 1970; 102: 855.

Figure 52-3 Electron micrograph of *M pneumoniae*-infected human fetal trachea after 48 hours in organ culture. **M**, mycoplasma; **C**, cilia; **L**, lumen; **arrows**, tip attachment structure. Bar = 1 μm. From Collier, A.M.: In: *The Mycoplasmas* Vol 2. Barile, M.F., et al., (editors). New York: Academic Press. 1979.

branched mycelioid structures, hence the name mycoplasma (*myces*, a fungus; *plasma*, a form). In the case of spiroplasmas, the filaments are helical, usually 2–5 μm long and 0.1–0.2 μm thick. Mycoplasmas reproduce by binary fission, but cytoplasmic division frequently may lag behind genome replication, resulting in formation of multinuclear filaments.

A few *Mycoplasma* sp possess unique attachment organelles shaped as a tapered tip in *M pneumoniae* and a pear-shaped bleb in the avian *M gallisepticum*. These two mycoplasmas are pathogens of the respiratory tract, adhering to the respiratory epithelium primarily through the terminal attachment structures (Figure 52-3). Interestingly, these mycoplasmas exhibit gliding motility on liquid-covered surfaces. The direction of movement always is led by the terminal structure, again indicating its importance in attachment.

One of the most useful distinguishing features of mycoplasmas is their peculiar fried-egg colony shape, consisting of a central zone of growth embedded in the agar and a peripheral zone on its surface (Figure 52-4). Among the prokaryotes, only the wall-less bacterial L-forms produce a similar fried-egg colony.

Nutrition and Energy-Yielding Mechanisms

The mycoplasmas have limited biosynthetic abilities, probably reflecting their small genome and parasitic mode of life. Consequently, they require complex media

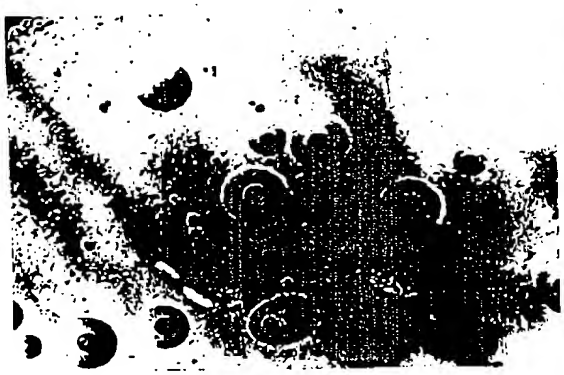

Figure 52-4 "Fried-egg" colonies of mycoplasmas. A 4-day culture on solid medium. From Razin, S., Oliver, O. *J Gen Microbiol* 1961; 24: 225.

containing serum for growth. The serum provides fatty acids and cholesterol for membrane synthesis in an assimilable, nontoxic form. The requirement of most mycoplasmas for cholesterol (Table 52-1) is unique among prokaryotes, although cholesterol can also be found in *Borrelia* cells and is apparently required for growth of *Treponema hyodysenteriae* (see Chapter 50). Cholesterol is incorporated into the cell membrane in large quantities and appears to function as a regulator of membrane fluidity. It must be stressed that the culture media now available are inadequate for cultivation of all mycoplasmas. Nevertheless, the continuing developments in culture media have allowed identification of a number of new species, including *Mycoplasma genitalium*, which was isolated from the human genital tract.

Glucose and other metabolizable carbohydrates are possible energy sources for the fermentative mycoplasmas possessing the Embden-Meyerhof-Parnas glycolytic pathway. All mycoplasmas examined so far possess a truncated flavin-terminated respiratory system, which rules out oxidative phosphorylation as an ATP-generating mechanism in mycoplasmas. Breakdown of arginine by the arginine dihydrolase pathway has been proposed as the major source of ATP in nonfermentative mycoplasmas. Ureaplasmas show the unique requirement among living organisms for urea. Because they are nonglycolytic and lack the arginine dihydrolase pathway, it has been suggested that ATP is generated in these organisms through an electrochemical gradient produced by the ammonia liberated during the intracellular hydrolysis of urea by the organism's urease.

Molecular Biology and Phylogenetic Status

The mycoplasma genome typically is prokaryotic, consisting of a circular, double-stranded DNA molecule. The *Mycoplasma* and *Ureaplasma* genomes are the smallest recorded for any self-reproducing prokaryote; the value of 23%–25% guanine plus cytosine (G + C) content found for the DNA of some mycoplasmas is also the lowest recorded (Table 52-1). Only one or two copies of ribosomal RNA genes exist in the mycoplasma genome, compared to seven in that of *E coli*, a finding that may explain the relatively slow growth rate of mycoplasmas. The protein synthesis machinery in

mycoplasmas is prokaryotic, so antibiotics such as chloramphenicol and tetracyclines inhibit protein synthesis and growth. Although it is too early to conclude whether mycoplasmas evolved from cell-wall-covered bacteria or vice versa, the repeated failure to show any genetic relationship between mycoplasmas and cell-wall-covered bacteria by DNA hybridization techniques contradicts the notion that mycoplasmas represent stable L-forms of existing cell-wall-covered prokaryotes.

The Cell Membrane

The lack of cell walls and intracytoplasmic membranes facilitates isolation of the mycoplasma membrane in a relatively pure form. The isolated mycoplasma membrane resembles that of other prokaryotes in being composed of approximately two-thirds protein and one-third lipid. The mycoplasma lipids resemble those of other bacteria, apart from the large quantities of cholesterol in the sterol-requiring mycoplasmas.

Membrane proteins, glycolipids, and lipoglycans exposed on the cell surface are the major antigenic determinants in mycoplasmas. Antisera containing antibodies to these components inhibit growth and metabolism of the mycoplasmas and, in the presence of complement, also cause lysis of the organisms. These properties are utilized in various serologic tests that differentiate between mycoplasma species and serotypes and detect antibodies to mycoplasmas in sera of patients (see Diagnosis section, page 615).

Some mycoplasmas are covered by capsules. In *M mycoides*, the contagious bovine pleuropneumonia agent, the capsule is made of galactan, a polymer possessing toxic properties. Electron microscopy has demonstrated capsules stained by ruthenium red in various mycoplasma species pathogenic to humans and animals. The possibility that capsules play a role in pathogenesis, by inhibiting phagocytosis of the organisms or by facilitating their adherence to host cell surfaces, remains to be studied.

Pathogenesis

Mycoplasmal Pneumonia

The term primary atypical pneumonia was coined in the early 1940s to describe pneumonias different from the typical lobar pneumonia caused by pneumococci. Several common respiratory viruses, including influenza and adenoviruses, were shown to be responsible for a significant number of these pneumonias. From other cases, many of which developed antibodies agglutinating red blood cells in the cold (cold agglutinins), an unidentified filterable agent was isolated by Eaton and associates and, accordingly, named Eaton agent. Identification of this agent as a new mycoplasma species was achieved in 1962, after its successful cultivation on cell-free media by Chanock, Hayflick, and Barile. This mycoplasma, named *M pneumoniae*, was the first clearly documented mycoplasma pathogenic for humans.

The effects of *M pneumoniae* on humans range from subclinical infection to upper respiratory disease to bronchopneumonia. The majority of human infections do not progress to a clinically evident pneumonia. When pneumonia occurs, the onset generally is gradual, and the clinical picture is one of a mild to moderately severe illness, with early

specific receptors for *M pneumoniae* on host cells. Monoclonal antibodies to one of these proteins, protein P1 (mol. wt. 165,000), inhibit attachment of the parasite. Ferritin labeling of the antibodies has shown that P1 concentrates on the tip structure of the mycoplasma, a finding that provides further support for the theory that the tip structure serves as an attachment organelle.

Tracheal organ cultures from the human fetus or from the Syrian hamster have been extensively used as experimental models to study the interaction of *M pneumoniae* with respiratory epithelium. Reduction of ciliary activity followed by complete ciliostasis is the most pronounced manifestation of injury to the tracheal or oviduct explants by mycoplasmas. As the infection progresses, the cilia are distorted and lost, and the superficial epithelial cells undergo desquamation. The histologic manifestations of tissue damage reflect pronounced alterations in the metabolism of the infected organ cultures, as reflected by decreased oxygen consumption and by decreased RNA and protein synthesis.

The nature of the toxic factors damaging the organ explants infected by mycoplasmas is still unclear. Toxins are rarely found in mycoplasmas. Consequently, researchers considered the possibility that the end products of mycoplasma metabolism may be responsible for tissue damage. Hydrogen peroxide, the end product of respiration in mycoplasmas, has been incriminated as a major pathogenic factor ever since it was shown to be responsible for the lysis of erythrocytes by mycoplasmas in vitro; however, the production of H_2O_2 alone does not determine pathogenicity, as the loss of virulence in *M pneumoniae* is not accompanied by a decrease in H_2O_2 production. For the H_2O_2 to exert its toxic effect, the mycoplasmas must adhere close enough to the host cell surface to maintain a toxic, steady-state concentration of H_2O_2 sufficient to cause direct damage, such as lipid peroxidation, to the cell membrane. The accumulation of malonyldialdehyde, an oxidation product of membrane lipids, in cells exposed to *M pneumoniae* supports this notion. Moreover, *M pneumoniae* has been found to inhibit host cell catalase. This would be expected to cause the accumulation of H_2O_2 at the site of parasite-host cell contact.

Ammonia is another end product of mycoplasma metabolism that may become toxic when produced in large quantities, as during urea hydrolysis by ureaplasmas. Hydrolysis of arginine by mycoplasmas possessing the arginine dihydrolase pathway also yields ammonia as an end product. In this case, however, the depletion of the essential amino acid, rather than the toxicity of the end product, is to blame for many of the symptoms in cell cultures contaminated by arginine-splitting mycoplasmas. Obviously, arginine depletion is unlikely to occur in the infected animal.

Evidence suggests that in *M pneumoniae* pneumonia, both organism-related and host-related factors are involved in pathogenesis. The host may be largely responsible for the appearance of pneumonia by mounting a local immunocyte and phagocytic response to the parasite. Syrian hamsters inoculated intranasally by *M pneumoniae* show patchy bronchopneumonic lesions consisting of infiltration of mononuclear cells. The ablation of thymic function before the experimental infection prevents development of the characteristic pulmonary infiltration, but lengthens the period during which the organisms can be isolated from the lung. Allowing the animals to recover and then reinfecting them produces an exaggerated and accelerated pneumonic process. Epidemiologic data also can be interpreted to show that repeated infections in humans are required before symptomatic disease occurs. Thus, serum antibodies to *M pneumoniae*

can be found in most children 2–5 years of age, although the illness occurs with greatest frequency in individuals 5–15 years of age.

An immunopathologic mechanism also may explain the complications affecting organs distant from the respiratory tract in some *M pneumoniae* patients. Various autoantibodies have been detected in the serum of many of these patients, including cold agglutinins reacting on the erythrocyte I antigen, antibodies reacting with lymphocytes, smooth muscle antibodies, and antibodies reacting with brain and lung antigens. Serologic cross-reactions between *M pneumoniae* and brain and lung antigens have been demonstrated, and these antigens probably are related to the glycolipids of *M pneumoniae* membranes, which are also found in most plants and in many bacteria. Clearly, host reaction varies markedly, as only about one-half the patients develop cold agglutinins, and complications are rare, even among individuals with antitissue globulins.

Host Defenses

Infection with *M pneumoniae* induces development of antibodies in the serum that fix complement, inhibit growth of the organism, and lyse the organism in the presence of complement. Generally, the first antibodies produced are of the IgM class, whereas later in convalescence the predominant antibody is IgG. Secretory IgA antibodies also develop and appear to play an important role in host resistance. The first infection in infancy usually is asymptomatic and generates a brief serum antibody response. Recurrent infections, which occur at approximately 2–4 year intervals, generate a more prolonged systemic antibody response and increasing numbers of circulating antigen-responsive lymphocytes. By late childhood, clinically apparent lower respiratory disease, including pneumonia, becomes more common. Thus, mycoplasma respiratory disease manifestations appear to vary, depending on the state of local and systemic immunity at the time of reinfection. One hypothesis holds that local immunity mediates resistance to infection and that systemic immunity contributes substantially to the pulmonary and systemic reactions characteristic of *M pneumoniae* pneumonia.

The relative importance of humoral and cell-mediated immunity in resistance to mycoplasma infections of the respiratory tract is still unclear. For many mycoplasma infections, such as bovine pleuropneumonia, resistance can be transferred with convalescent serum. Although these results indicate that antibody can mediate resistance to mycoplasma infections of the respiratory tract, this may not be true for all mycoplasma respiratory diseases. Thus, resistance of rats to pulmonary disease induced by *M pulmonis* can be transferred only with spleen cells obtained from previously infected animals. Although IgA antibody may be important in upper respiratory tract resistance to mycoplasmas, other factors seem to be involved in resistance to pulmonary disease and these factors may not be the same for all mycoplasma infections.

Epidemiology

One of the most puzzling features of *M pneumoniae* pneumonia is the age distribution of patients. In a survey conducted between 1964 and 1975 of more than 100,000 individuals in the Seattle area, the age-specific attack rate was highest among the 5–9 year old

children. Rates of *M pneumoniae* pneumonia in the youngest age group, 0–4 years old, were about one-half those of school-age children, but considerably higher than among adults. *M pneumoniae* pneumonia was rarely observed in infants under the age of 6 months, suggesting maternally conferred immunity. *M pneumoniae* accounts for as much as 8%–15% of all pneumonias in young school-aged children. In older children and young adults, the organism is responsible for approximately 15%–50% of all pneumonias. Infection with *M pneumoniae* is worldwide and endemic and occurs all year round but shows a predilection for the colder months, apparently because of greater opportunity for transmission by droplet infection. *M pneumoniae* appears to require close personal contact to spread; successful spreading usually occurs in families, schools, and institutions. The incubation period is relatively long, ranging from 2–3 weeks.

U urealyticum is acquired primarily through sexual contact. Colonization has been linked to the frequency of sexual intercourse and the number of sexual partners. Women may serve as asymptomatic reservoirs of infection, so simultaneous treatment of both sexual partners is necessary to cure men suffering from nongonococcal urethritis caused by *U urealyticum*.

Diagnosis

Diagnosis on the basis of microscopy alone is equivocal; therefore, cultivation is essential for definitive diagnosis.

Cultivation

A routine mycoplasma medium consists of heart infusion, peptone, yeast extract, salts, glucose or arginine, and horse serum (5%–20%). Fetal or newborn calf sera are preferable to horse serum. To prevent the overgrowth of the fast-growing bacteria that usually accompany mycoplasmas in clinical materials, penicillin or thallium acetate or both are added as selective agents. To cultivate ureaplasmas, the medium is supplemented with urea, and its pH is brought to 6.0. Ureaplasmas and *M genitalium* are relatively sensitive to thallium, so this inhibitor is omitted from their culture media. To isolate *M pneumoniae*, nasopharyngeal secretions are inoculated into a selective diphasic medium (pH 7.8) made of mycoplasma broth and agar and supplemented with glucose and phenol red. When *M pneumoniae* grows in this medium, it produces acid, causing the color of the medium to change from purple to yellow. Broth from the diphasic medium is subcultured to mycoplasma agar when a color change occurs, or at weekly intervals for a minimum of 8 weeks.

Identification

Colonies appearing on the plates can be identified as *M pneumoniae* by staining directly on agar with homologous fluorescein-conjugated antibody, or by demonstrating that a specific antiserum to *M pneumoniae* inhibits their growth on agar. Colonies of ureaplasmas are usually minute (less than 100 μm in diameter); because of urea hydrolysis and ammonia liberation, the pH of the medium becomes alkaline. When manganous sulfate is added to the growth medium, the colonies of ureaplasmas stain

dark brown. More detailed characterization of isolates can be achieved by a variety of routine biochemical and serologic tests, supplemented when required by more sophisticated tests, including electrophoretic analysis of cell proteins, crossed immunoelectrophoresis of cell proteins, DNA hybridization tests, and DNA cleavage pattern evaluations by use of restriction endonucleases.

Serodiagnosis

Serodiagnosis basically consists of demonstrating antibodies in the patients' sera that inhibit growth and metabolism of the organism or fix complement with mycoplasmal antigens. Antibody response in mycoplasmal pneumonia is most easily demonstrated by complement fixation, reacting acute- and convalescent-phase sera with intact organisms or their lipid extract as antigen. A fourfold or greater antibody rise is considered significant, because a relatively high level of antibody may persist for 1 or more years after infection. The cold agglutinin test is less useful because only about one-half of patients develop cold agglutinins, and because these antibodies also are induced by a great many other conditions.

Present techniques for laboratory diagnosis of *M pneumoniae* infections are of little use to the clinician because recovery and identification of the mycoplasmas cannot be accomplished in less than 1–2 weeks. Development of methods for rapid laboratory diagnosis, such as direct demonstration of organisms in the sputum by immunofluorescence, electron microscopy, or the ELISA technique, will be of great value.

Control

Prevention

Chemoprophylaxis of mycoplasma infections is not recommended, because it does not cure the infection, although it may modify the secondary cases to subclinical disease. Attempts at control by immunoprophylaxis fail in most cases. Prior natural infection appears to provide the most effective resistance; however, evidence shows that *M pneumoniae* infections in humans recur at intervals of several years. These observations suggest that immunity to a single natural infection is relatively short-term, particularly in children, and it may be unrealistic to expect more, or even as much, from artificially induced immunity.

Attenuation of mycoplasma strains tends to reduce virulence and immunogenicity. In most cases, attenuated viable vaccines do not reach the level of protective efficiency required from a commercial vaccine. *M mycoides* is an exception, because subcutaneous inoculation of live organisms in the tail protects cattle effectively against contagious bovine pleuropneumonia. Killed *M pneumoniae* vaccines administered intranasally to hamsters are relatively ineffective unless boosted by parenteral inoculation of vaccine. Intranasal immunizations may be ineffective because the antigenic mass is not retained for a sufficient time in the lung. On the other hand, parenteral killed vaccines, particularly if combined with adjuvant, do produce adequate protection by reducing

pneumonia, although a minimal effect on the number of organisms growing in the lungs can be demonstrated. A similar protective effect can be achieved briefly by inoculation of hyperimmune serum.

In summary, a single dose of vaccine in a form suitable for clinical use is unlikely to produce lasting immunity to mycoplasma infection. Stimulation of systemic antibodies may prevent the clinical manifestations of pneumonia, but additional local stimulation with live or killed organisms may be necessary to evoke resistance to colonization. An approach worth pursuing is the preparation of vaccines made of antigenic components specifically related to the mycoplasma-host cell interaction, such as components of the mycoplasma membrane responsible for attachment of the parasites to the epithelial cell surface.

Treatment

The mycoplasmas are sensitive to most broad-spectrum antibiotics such as tetracyclines and chloramphenicol but are resistant to antibiotics that specifically inhibit bacterial cell wall synthesis. Tetracycline combined with erythromycin therapy has been definitely shown to reduce duration of fever and pulmonary infiltrations in *M pneumoniae* patients. Effective treatment of the disease symptoms, however, usually is not accompanied by eradication of the organism from the infected host. To prevent recurrence of nongonococcal urethritis caused by *U urealyticum*, sex partners should be treated simultaneously with tetracyclines.

r nc s

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(54) Title: ANTIGEN COMPOSITION AGAINST MYCOPLASMA

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1234567890 1234567890 1234567890 1234567890 1234567890

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HKVSNDSIRI ALTDPDNPRW ISAQKDIISY VDETEAATST ITKNQDAQNN     100
WLTQQANLSP APKGFIIAPE NGSGVGTAVN TIADKGIPIV AYDRLITGSD     150
KYDWYVSFDN EKVGELQGLS LAAGLLGKED GAFDSIDQMN EYLKSHMPQE     200
TISFYTIAGS QDDNNSQYFY NGAMKVLKEL MKNSQNKIID LSPEGENAVY     250
VPGWNYGTAG QRIQSFLTIN KDPAGGNKIK AVGSKPASIF KGFLAPNDGM     300
AEQAITKLKL EGFDTQKIFV TRQDYNDKAK TFIKDGDQNM TIYKPDKVLG     350
KVAVEVLRVL IAKKNKASRS EVENELKAKL PNISFKYDNQ TYKVQGKNIN     400
TILVSPVIVT KANVDNPDA                                       419
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(57) Abstract

The present invention relates to a putative protective antigen against a *Mycoplasma*, prepared by a method including a sample of a *Mycoplasma*; an antibody probe including at least one antibody against a *Mycoplasma* produced by a method including: providing a biological sample taken a short time after an immune animal has been challenged with a *Mycoplasma* or *Mycoplasma* extract taken from the infection site or an area of a lesion or an area close to the infection site or lesion; isolating cells from the biological sample; culturing cells *in vitro* in a suitable culture medium; and harvesting antibodies produced from said cells; probing the *Mycoplasma* sample with the antibody probe to detect at least one antigen; and isolating the antigen detected, also including diagnostic antigens, the preparation thereof, and their use in the formation of vaccine compositions, particularly vaccine compositions against *Mycoplasma hyopneumoniae* infections.

ANTIGEN COMPOSITION AGAINST MYCOPLASMA

The present invention relates to protective and diagnostic antigens, the preparation thereof, and their use in the formation of vaccine compositions, particularly vaccine compositions against Mycoplasma hyopneumoniae infections.

Mycoplasma hyopneumoniae is a ubiquitous swine respiratory pathogen causing mycoplasmal pneumoniae in swine (swine enzootic pneumonia). Swine enzootic pneumonia is probably the most widespread and economically significant disease in swine producing countries of the world. The economic effects of swine enzootic pneumonia (SEP) are complex, and the cost of the disease is severe. In Australia, the disease was estimated in 1988 to cost approximately $20,000,000 per annum. Increased mortality, decreased growth weight, depressed feed conversion, susceptibility to secondary bacterial infections, increased management costs, and increased use of antibiotics, are the main reasons for the economic impact of SEP.

Whilst several experimental vaccines have been produced, these have resulted in less than optimal results, and utilising various classes of antibiotics such as tetracycline, lincamycin and tiamulin is still the most widespread control treatment. Such antibiotics are, however, of limited therapeutic value, because they do not prevent the establishment of an infection, and lung lesions may develop after treatment ends.

European Patent Application 359,919 to ML Technology Ventures L.P. describes a series of antigens, 36 kD, 41 kD, 74.5 kD and 96 kD in size, and proposes the use of such antigens in vaccines. Results presented suggest that some protection in pigs against challenge was achieved.

However, there remains a need in the art for an effective vaccine against M. hyopneumoniae which would confer protection against colonisation and clinical disease following M. hyopneumoniae challenge and also significantly reduce the morbidity and mortality from secondary infections.

Accordingly, it is an object of the present invention to overcome, or at least alleviate, one or more of the difficulties and deficiencies in the prior art.

Accordingly, in a first aspect of the present invention there is provided a putative protective antigen against a Mycoplasma, preferably Mycoplasma

hyopneumoniae prepared by a method including

providing

a sample of a Mycoplasma;

an antibody probe including at least one antibody against a Mycoplasma produced by a method including;

providing a biological sample taken a short time after an immune animal has been challenged with a Mycoplasma or Mycoplasma extract taken from the infection site or an area of a lesion or an area close to the infection site or lesion;

isolating cells from the biological sample;

culturing cells in vitro in a suitable culture medium; and

harvesting antibodies produced from said cells;

probing the Mycoplasma sample with the antibody probe to detect at least one antigen; and

isolating the antigen detected.

The protective antigens may also function as diagnostic antigens as discussed below.

Accordingly, in a preferred aspect of the present invention there is provided a putative protective antigen against Mycoplasma hyopneumoniae, or related infections, selected from the group of antigens having approximate molecular weights of 110-114, 90-94, 72-75, 60-64, 52-54 and 46-48 kilodaltons (kD), as hereinafter described, mutants, derivatives and fragments thereof. The putative protective antigen may be a surface protein. The putative protective antigen may be a surface lipoprotein or membrane protein.

Preferably the protective antigens are selected from the group of antigens having approximate molecular weights of 110-114, 90-94, 74, 62, 52 and 48 kD.

Preferably, the 72-75 kD antigen includes the following N-terminal amino acid sequence:

AGXLQKNSLLEEVWYLAL

and, optionally, one or more of the following internal amino acid sequences:

AKNFDFAPSIQGYKKIAHEL

NLKPEQILQLLG

LLKAEXNKXIEEINTXLDN

Preferably, the 60-64 kD antigen includes one of the following N-terminal amino acid sequences:

MKLAKLLKGFX(N/L)(M/V)IK

ADP(F/I)(R/E)Y(V/A)PQG(Q/A)X(M/N)VG

Preferably, the 52-54 kD antigen includes the following N-terminal amino acid sequence:

AGXWAKETTKEEKS

and, optionally, one or more of the following internal amino acid sequences:

AWVTADGTVN

AIVTADGTVNDNKPNQWVRKY.

Preferably, the 46-48 kD antigen includes the following N-terminal amino acid sequence:

AGXGQTESGSTSDSKPQAETLKHKV

and, optionally, one or more of the following internal amino acid sequences:

TIYKPDKVLGKVAVEVLRVLIAKKNKASR

AEQAITKLKLEGFDTQ

KNSQNKIIDLSPEG

The 46-48 kD antigen may be encoded by a nucleic acid fragment:

10 1234567890	20 1234567890	30 1234567890	40 1234567890	50 1234567890	
ATGAAAAAAA	TGCCACTATA	CCAGAGGAAA	GAGCAGTATA	TAAAATAATT	50
AAAATTACAT	TTTCTTCATT	TGCGCCAGAA	TTTTTAAGAA	TTAGTACATT	100
AAAAAGTAGA	ACAAAAGTTA	TTAATGTAAA	CATTAGCGCA	ATCCTTAAGA	150
AAAAATTAAA	AGTTTTATCT	ATTTTTTTTA	ATCGAAATCC	AACCAGGCAT	200
AAATCTTTGT	CAGTATTTAT	CAAGTCGGTA	TTTTTTCATT	ATTTCTACTA	250
AAATATTATT	TGAATTTGCA	TTTTCCATAA	TCTAAAATTT	TACATTTTTT	300
TATAACAATT	TTTAAAAATT	ACTCTTTAAT	TTATAGTATT	TTTTTATTTT	350
TTAGTCTAAA	TTATAAAATT	ATCTTGAATT	TTATTTGAAT	TTTTATAATT	400
TAGTACTAAA	AAATACAAAT	ATTTTTTCCT	ATTCTAAGAA	AAATTCATTT	450
TTTAAAAAAA	ATTGATTTTT	ATAGTATAAT	TTGTTTGTAT	AATTGAATTA	500
ACTTGATTTG	AAAGGGAACA	AAATGAAAAA	AATGCTTAGA	AAAAAATTCT	550
TGTATTCATC	AGCTATTTAT	GCAACTTCGC	TTGCATCAAT	TATTGCATTT	600
GTTGCAGCAG	GTTGTGGACA	GACAGAATCA	GGTTCAACTT	CTGATTCTAA	650
ACCACAAGCC	GAGACGCTAA	AACATAAAGT	AAGTAATGAT	TCTATTCGAA	700
TAGCACTAAC	CGATCCGGAT	AATCCTCGAT	GAATTAGTGC	CCAAAAAGAT	750
ATTATTTCTT	ATGTTGATGA	AACAGAGGCA	GCAACTTCAA	CAATTACAAA	800
AAACCAGGAT	GCACAAAATA	ACTGACTCAC	TCAGCAAGCT	AATTTAAGCC	850
CAGCGCCAAA	AGGATTTATT	ATTGCCCCTG	AAAATGGAAG	TGGAGTTGGA	900

ACTGCTGTTA	ATACAATTGC	TGATAAAGGA	ATTCCGATTG	TTGCCTATGA	950
TCGACTAATT	ACTGGATCTG	ATAAATATGA	TTGGTATGTT	TCTTTTGATA	1000
ATGAAAAAGT	TGGTGAATTA	CAAGGTCTTT	CACTTGCTGC	GGGTCTATTA	1050
GGAAAAGAAG	ATGGTGCTTT	TGATTCAATT	GATCAAATGA	ATGAATATCT	1100
AAAATCACAT	ATGCCCCAAG	AGACAATTTC	TTTTTATACA	ATCGCGGGTT	1150
CCCAAGATGA	TAATAATTCC	CAATATTTTT	ATAATGGTGC	AATGAAAGTA	1200
CTTAAAGAAT	TAATGAAAAA	TTCGCAAAAT	AAAATAATTG	ATTTATCTCC	1250
TGAAGGCGAA	AATGCTGTTT	ATGTCCCAGG	ATGAAATTAT	GGAACTGCCG	1300
GTCAAAGAAT	CCAATCTTTT	CTAACAATTA	ACAAAGATCC	AGCAGGTGGT	1350
AATAAAATCA	AAGCTGTTGG	TTCAAAACCA	GCTTCTATTT	TCAAAGGATT	1400
TCTTGCCCCA	AATGATGGAA	TGGCCGAACA	AGCAATCACC	AAATTAAAAC	1450
TTGAAGGGTT	TGATACCCAA	AAAATCTTTG	TAACTCGTCA	AGATTATAAT	1500
GATAAAGCCA	AAACTTTTAT	CAAAGACGGC	GATCAAAATA	TGACAATTTA	1550
TAAACCTGAT	AAAGTTTTAG	GAAAAGTTGC	TGTTGAAGTT	CTTCGGGTTT	1600
TAATTGCAAA	GAAAAATAAA	GCATCTAGAT	CAGAAGTCGA	AAACGAACTA	1650
AAAGCAAAAC	TACCAAATAT	TTCATTTAAA	TATGATAATC	AAACATATAA	1700
AGTACAAGGT	AAAAATATTA	ATACAATTTT	AGTAAGTCCA	GTAATTGTTA	1750
CAAAAGCTAA	TGTTGATAAT	CCTGATGCCT	AA		1782

Accordingly, in a further aspect the present invention provides an isolated nucleic acid fragment encoding a putative protective antigen against Mycoplasma hyopneumoniae or related infections, said nucleic acid fragment:

10	20	30	40	50	
1234567890	1234567890	1234567890	1234567890	1234567890	
ATGAAAAAAA	TGCCACTATA	CCAGAGGAAA	GAGCAGTATA	TAAAATAATT	50
AAAATTACAT	TTTCTTCATT	TGCGCCAGAA	TTTTTAAGAA	TTAGTACATT	100
AAAAAGTAGA	ACAAAAGTTA	TTAATGTAAA	CATTAGCGCA	ATCCTTAAGA	150
AAAAATTAAA	AGTTTTATCT	ATTTTTTTTA	ATCGAAATCC	AACCAGGCAT	200
AAATCTTTGT	CAGTATTTAT	CAAGTCGGTA	TTTTTTCATT	ATTTCTACTA	250
AAATATTATT	TGAATTTGCA	TTTTCCATAA	TCTAAAATTT	TACATTTTTT	300
TATAACAATT	TTTAAAAATT	ACTCTTTAAT	TTATAGTATT	TTTTTATTTT	350
TTAGTCTAAA	TTATAAAATT	ATCTTGAATT	TTATTTGAAT	TTTTATAATT	400
TAGTACTAAA	AAATACAAAT	ATTTTTTCCT	ATTCTAAGAA	AAATTCATTT	450
TTTAAAAAAA	ATTGATTTTT	ATAGTATAAT	TTGTTTGTAT	AATTGAATTA	500
ACTTGATTTG	AAAGGGAACA	AAATGAAAAA	AATGCTTAGA	AAAAAATTCT	550
TGTATTCATC	AGCTATTTAT	GCAACTTCGC	TTGCATCAAT	TATTGCATTT	600
GTTGCAGCAG	GTTGTGGACA	GACAGAATCA	GGTTCAACTT	CTGATTCTAA	650
ACCACAAGCC	GAGACGCTAA	AACATAAAGT	AAGTAATGAT	TCTATTCGAA	700
TAGCACTAAC	CGATCCGGAT	AATCCTCGAT	GAATTAGTGC	CCAAAAAGAT	750
ATTATTTCTT	ATGTTGATGA	AACAGAGGCA	GCAACTTCAA	CAATTACAAA	800
AAACCAGGAT	GCACAAAATA	ACTGACTCAC	TCAGCAAGCT	AATTTAAGCC	850
CAGCGCCAAA	AGGATTTATT	ATTGCCCCTG	AAAATGGAAG	TGGAGTTGGA	900
ACTGCTGTTA	ATACAATTGC	TGATAAAGGA	ATTCCGATTG	TTGCCTATGA	950
TCGACTAATT	ACTGGATCTG	ATAAATATGA	TTGGTATGTT	TCTTTTGATA	1000
ATGAAAAAGT	TGGTGAATTA	CAAGGTCTTT	CACTTGCTGC	GGGTCTATTA	1050
GGAAAAGAAG	ATGGTGCTTT	TGATTCAATT	GATCAAATGA	ATGAATATCT	1100
AAAATCACAT	ATGCCCCAAG	AGACAATTTC	TTTTTATACA	ATCGCGGGTT	1150
CCCAAGATGA	TAATAATTCC	CAATATTTTT	ATAATGGTGC	AATGAAAGTA	1200
CTTAAAGAAT	TAATGAAAAA	TTCGCAAAAT	AAAATAATTG	ATTTATCTCC	1250
TGAAGGCGAA	AATGCTGTTT	ATGTCCCAGG	ATGAAATTAT	GGAACTGCCG	1300
GTCAAAGAAT	CCAATCTTTT	CTAACAATTA	ACAAAGATCC	AGCAGGTGGT	1350

AATAAAATCA	AAGCTGTTGG	TTCAAAACCA	GCTTCTATTT	TCAAAGGATT	1400
TCTTGCCCCA	AATGATGGAA	TGGCCGAACA	AGCAATCACC	AAATTAAAAC	1450
TTGAAGGGTT	TGATACCCAA	AAAATCTTTG	TAACTCGTCA	AGATTATAAT	1500
GATAAAGCCA	AAACTTTTAT	CAAAGACGGC	GATCAAAATA	TGACAATTTA	1550
TAAACCTGAT	AAAGTTTTAG	GAAAAGTTGC	TGTTGAAGTT	CTTCGGGTTT	1600
TAATTGCAAA	GAAAAATAAA	GCATCTAGAT	CAGAAGTCGA	AAACGAACTA	1650
AAAGCAAAAC	TACCAAATAT	TTCATTTAAA	TATGATAATC	AAACATATAA	1700
AGTACAAGGT	AAAAATATTA	ATACAATTTT	AGTAAGTCCA	GTAATTGTTA	1750
CAAAAGCTAA	TGTTGATAAT	CCTGATGCCT	AA		1782

As cross protection between various Mycoplasma such as M. hyorhinis and M. synoviae has been documented, similar antigens may also be detected in other Mycoplasma species (Figure 1).

In a still further aspect the present invention provides a method for preventing Mycoplasma infection in animals. Preferably the Mycoplasma disease is a Mycoplasma hyopneumoniae disease such as swine enzootic pneumonia (SEP). This method includes administering to an animal an effective amount of at least one protective antigen against Mycoplasma as described above.

The present invention further provides a vaccine composition including a prophylactically effective amount of at least one putative protective antigen against a Mycoplasma as herein described. Preferably the veterinary composition includes two or more putative protective antigens as herein described.

Accordingly in a preferred aspect the present invention provides a vaccine composition including two or more putative protective antigens selected from antigens having approximate molecular weights of 110-114, 90-94, 72-75, 60-64, 52-54 and 46-48 kilodaltons.

The vaccine composition may include any combination of two or more putative protective antigens selected from antigens having approximate molecular weights of 110-114, 90-94, 72-75, 60-64, 52-54 and 46-48 kD. The two or more antigens may be selected from antigens falling within one of the specified approximate molecular weights and/or antigens from different specified approximate molecular weights. The composition may contain 3, 4, 5 or 6 antigens selected from protective antigens having molecular weights of approximately 110-114, 90-94, 72-75, 60-64, 52-54 and 46-48 kD.

The vaccine compositions according to the present invention may be

administered orally or may be administered parenterally (for example by intramuscular, subcutaneous, intradermal or intravenous injection). The amount required will vary with the antigenicity of the active ingredient and need only be an amount sufficient to induce an immune response typical of existing vaccines.

Reactive experimentation will easily establish the required amount. Typical initial doses of vaccine or veterinary compositions may be approximately 0.001-1 mg active ingredient/kg body weight. The dose rate may increase or multiple doses may be used as needed to provide the desired level of protection.

The vaccine composition according to the present invention may further include a veterinary acceptable carrier, diluent or excipient therefor. Preferably the active ingredient may be suspended or dissolved in a carrier. The carrier may be any solid or solvent that is nontoxic to the animal and compatible with the active ingredient. Suitable carriers include liquid carriers, such as normal saline and other nontoxic salts at or near phsyiological concentrations, and solid carriers, such as talc or sucrose.

Preferably the vaccine contains an adjuvant, such as Freund's adjuvant, complete or incomplete, or immunomodulators such as cytokines may be added to enhance the antigenicity of the antigen if desired.

More preferably the adjuvant is of the mineral-oil type as these have been found to be consistently superior at inducing antibody titres and Delayed Type Hypersensitivity responses. A particularly preferred adjuvant is that marketed under the trade designation Montanide ISA-50 and available from Seppic, Paris, France.

When used for administering via the bronchial tubes, the vaccine is suitably present in the form of an aerosol.

In a still further aspect of the present invention there is provided a diagnostic kit including a diagnostic antigen against a Mycoplasma, preferably Mycoplasma hypneumoniae, identified and purified as described above.

The putative protective antigens according to the present invention may be isolated and identified utilising the general methods described in Australian patent application 49035/90, the entire disclosure of which is incorporated herein by reference.

Accordingly, in a further aspect, the present invention provides a method for producing at least one antibody against a Mycoplasma. This method includes

providing a biological sample taken a short time after an immune animal has been challenged with a Mycoplasma or Mycoplasma extract taken from the infection site or an area of a lesion or an area close to the infection site or lesion;

isolating cells from the biological sample;

culturing cells in vitro in a suitable culture medium; and

harvesting antibodies produced from said cells.

The Mycoplasma may be Mycoplasma hyopneumoniae.

The animal may be a mammal including humans. The mammal may be a domestic animal such as a pig, sheep or cattle.

The biological animal sample may be of any suitable type. The biological sample may be taken from animal tissue, organs, lymph or lymph nodes. The biological sample may be taken from the infection site, the lungs of the animal, or an area of a lesion which may be formed or an area close to the infected site or a lesion such as in the lymph nodes draining from the lungs.

However, serum/plasma samples are not used as the biological samples according to this aspect of the present invention. It has been found that the majority of antibodies found in a serum/plasma sample are irrelevant to protection or specific diagnosis or a Mycoplasma or are unrelated to the Mycoplasma. In addition, other serum/ plasma components may interfere with the specific reactions between pathogen components and antibodies to them.

In contrast, the probes described in the present invention are highly enriched in Mycoplasma-specific antibodies of particular importance to protective immunity.

It is preferred that the biological samples are taken from the animals at a predetermined time in the development of the disease. In general, for a Mycoplasma infection, it has been found that the biological samples should be taken approximately 2 to 7 days after challenge with or after administration of products obtained from a pathogen or with the pathogen itself.

The cells isolated from the biological sample may include B cells.

Thus, preferably the cells are taken a short time after in vivo stimulation,

preferably within approximately 2 to 5 days thereafter, resulting in the in vivo induction of antibody forming cells which will secrete specific antibodies into the culture medium after in vitro incubation.

In vitro secretion of antibodies in the culture medium by recently activated B cells may be enhanced by the addition of helper factors to the cultures. The helper factors may be cytokines used alone or in combination, including Interleukin 1, 2, 3, 4, 5, 6, 7 and 8, colony stimulating factors, interferons and any other factors that may be shown to have an enhancing effect on specific B cell secretion.

The method of producing an antibody may include a further step of activating the cells isolated to proliferate and secrete and/or release antibodies.

The cell activation step may include adding a cell activating agent to the culture medium. The cell activating agent may be selected from mitogens and helper factors produced by leukocytes, or their synthetic equivalents or combinations thereof.

The mitogens may be selected from the group including products derived from pokeweed (Phytolacca americana) also known as pokeweed mitogen (PWM), polyvinylpyrrolidone (PVP), polyadenylic-polyuridylic acid (poly(A-U)), purified protein derivate (PPD), polyinosinic-polycytidilic acid (poly(I-C)), lipopolysaccharide (LPS), staphylococcal organisms or products thereof, Bacto-streptolysin O reagent (SLO), Staphylococcal phage lysate (SPL), Epstein-Barr virus (EBV), Nocardia water-soluble mitogen (NWEM), phytohemagglutinin (PHA), Concanavalin A (Con A), and dextran-sulphate and mixtures thereof. The cell proliferation agent may be any agent that indirectly or directly results in B cell proliferation and/or antibody secretion such as solid-phase anti-immunoglobulin. The helper factors may be selected from the group including cytokines including interleukin 1, 2, 3, 4, 5, 6, 7 and 8, colony stimulating factors, interferons and any other helper factors that may be shown when added alone, or in combination with other factors and agents, to have an enhancing effect on specific B cell proliferation and/or antibody secretion. This in no way is meant to be an exhaustive list of mitogens and cell actuating agents including helper factors.

The in vitro culturing of the cells may be conducted with or without prior

steps to separate sub-populations of cells. The harvesting of antibodies may be conducted by harvesting of the supernatant from the culture medium. This supernatant contains antibodies secreted by these cells during the in vitro culture or artificially released from the B cells, for example by lysis of the B cells. It has been found that the antibody-containing supernatants may be used directly to detect antigens of the Mycoplasma.

In a preferred aspect of the present invention, there is provided a method for identifying an antigen associated with a Mycoplasma, preferably Mycoplasma hyopneumoniae. This method includes

providing

a sample of a Mycoplasma; and

an antibody probe including at least one antibody against a Mycoplasma;

probing the Mycoplasma sample with the antibody probe to detect at least one antigen; and

isolating the antigen detected.

The sample of Mycoplasma may be mixed with a standard buffer solution and placed on a standard support such as an SDS-polyacrylamide gel to separate the proteins contained thereon (Figure 2).

Alternatively, the proteins may be selected utilising the non-ionic detergent Triton X-114 (TX-114). Insoluble material may be removed by centrifugation. Proteins soluble in the TX-114 phase may then be precipitated out (Figure 2).

The separate proteins may then be transferred to nitrocellulose, nylon or other sheets.

The probing with a suitable antibody may further include subjecting the product produced thereby to a detection assay. The detection assay may include Western blot techniques. The detection assay may be an immunoprecipitation assay, a radioimmunoassay, an enzyme-linked immunoassay or immunofluorescent assay (Figures 3, 4 and 5).

The antibody produced as described above may be utilized simply in the form of the supernatant harvested from the culture medium. Alternatively, the antibodies may be separated and purified.

In a further preferred aspect of the present invention the antibody contained in the culture medium may be used for the affinity purification, preferably immuno-affinity purification of antigen.

Accordingly, in a preferred aspect there is provided a method for purifying antigen. This method includes

providing

a crude antigen mixture; and

an antibody against a Mycoplasma immobilized on a suitable support;

subjecting the crude antigen mixture to affinity chromatography utilizing the immobilized antibody; and

isolating the purified antigen so formed.

The antibody is produced by the method described above.

Antibody can be obtained from the culture supernatant probe by conventional methods. For example, methods usually used to purify immunoglobulins from serum or plasma, e.g. precipitation with ammonium sulphate, fractionation with caprylic acid, ion exchange chromatography, or by binding and elution from immobilized protein G or protein A, may be utilized. Antibody so obtained can then be coupled to suitable supports, e.g., CNBr-activated Sepharose 4B (Pharmacia), Affi-gel (Bio-RAD), or other affinity chromatography supports able to bind proteins.

Immobilized antibody can then be applied to the fractionation and purification of specific antigen from a complex Mycoplasma extract by affinity chromatography. After binding of antigen to immobilized antibody, unbound macromolecular species can be washed away from the solid support with, e.g. buffers containing 1.5 M NaCl. Subsequently the antigen can be eluted from the affinity column with, e.g. low or high pH buffer or buffers containing chaotropic ions, e.g. 0.5-3.0 M sodium thiocyanate.

The application of the antibody probe to affinity chromatography enables sufficient quantities of specific antigens to be rapidly isolated from a complex crude extraction mixture for biochemical characterization, amino-acid sequencing and vaccination of animal for limited protection studies. Application of affinity

chromatography for obtaining antigen(s) avoids the difficulties often encountered when applying conventional biochemical techniques to the purification of an antigen about which little or no data is known. It also obviates the need to raise polyclonal or monoclonal antibodies for the purpose of "analytical" affinity chromatography. Large scale preparation may, however, require the preparation of polyclonal or monoclonal antibodies.

Having identified the antigen(s) molecular biology, chemical techniques, e.g. cloning techniques, may be used to produce unlimited amounts of this antigen or, alternatively, synthetic peptides corresponding to different fragments of the identified antigens may be used as a means to produce a vaccine.

Accordingly in a preferred aspect of the present invention there is provided a method for preparing a synthetic antigenic polypeptide against Mycoplasma, preferably Mycoplasma hypneumoniae, which method includes

providing

a cDNA library or genomic library derived from a sample of Mycoplasma; and

an antibody probe as described above;

generating synthetic polypeptides from the cDNA library or genomic library;

probing the synthetic polypeptides with the antibody probe; and

isolating the synthetic antigenic polypeptide detected thereby.

Either cDNA or genomic libraries may be used. The cDNA or genomic libraries may be assembled into suitable expression vectors that will enable transcription and the subsequent expression of the clone cDNA, either in prokaryotic hosts (e.g. bacteria) or eukaryotic hosts (e.g. mammalian cells). The probes may preferably be selected from

(i) synthetic oligonucleotide probes based on the amino acid sequence of the antigen identified and purified as described above;

(ii) antibodies obtained from the culture medium produced as described above;

(iii) monoclonal or polyclonal antibodies produced against the antigens identified and purified as described above;

(iv) recombinant or synthetic monoclonal antibodies or polypeptides with

specificity for the antigen, e.g. as described by Ward et al., Nature, 241, pages 544-546 (1989).

The synthetic antigenic polypeptide produced in accordance with the invention may be a fusion protein containing the synthetic antigenic peptide and another protein.

In a further aspect of the present invention there is provided a DNA fragment encoding a putative protective antigen against Mycoplasma or related infections, said DNA fragments having a nucleic acid sequence according to Figure 6a and 6b or an homologous sequence and functionally active fragments thereof.

In a further preferred aspect of the present invention there is provided a clone including a DNA fragment encoding a putative protective antigen against Mycoplasma or related infections, said DNA fragments having a nucleic acid sequence according to Figure 6a and 6b or an homologous sequence and functionally active fragments thereof.

Preferably the clone is pC1-2.

The present invention will now be more fully described with reference to the accompanying Examples and drawings. It should be understood, however, that the description following is illustrative only and should not be taken in any way as a restriction on the generality of the invention described above.

IN THE FIGURES:

FIGURE 1: SDS-Polyacrylamide gel (12.5%) profiles of SDS extracts of species of mycoplasma- Coomassie R250 stained.

Lane 1 Pre-stained Molecular Weight Standards.

Lane 2 *M. gallisepticum.*

Lane 3 *M. synoviae.*

Lane 4 *M. hyopneumoniae.*

Lane 5 *M. hyorhinis.*

Lane 6 *M. flocculare.*

FIGURE 2: SDS-Polyacrylamide gel (12.5%) profiles of extracts of strains *of M. hyopneumoniae* - Coomassie R250 stained gel

Lane 1 Pre-stained Molecular Weight Standards.

Lane 2 Triton X-114 extract of *M. hyopneumoniae* - strain Beaufort.

Lane 3 As for Lane 2.

Lane 4 SDS extract of *M. hyopneumoniae* strain Beaufort.

Lane 5 SDS extract of *M. hyopneumoniae* strain 10110.

FIGURE 3: Western blots of Triton X-114 extracted antigens from *M. hyopneumoniae* strain Beaufort, probed with serum and supernatant antibody probes.

Lane 1 No antibody control.

Lane 2 Dookie pig serum control 1/200.

Lane 3 Pig 105 supernatant.

Lane 4 Pig 1 supernatant.

Lane 5 Dookie pig supernatant.

FIGURE 4: Western blots of SDS extracted antigens from *M. hyopneumoniae* strain Beaufort probed with paired serum and supernatantantibody probes. Fractionation of antigens on SDS Polyacrylamide gel (12.5%).

Lane 1 a) Pig 453 supernatant.

b) Pig 453 serum 1/100.

Lane 2 a) Pig 105 supernanant.

b) Pig 105 serum 1/100.

Lane 3 a) Pig 1 superanatant.
b) Pig 1 serum 1/100.
Lane 4 a) Pig 15 supernanant.
b) Pig 15 serum 1/100.
Lane 5 a) Dookie supernatant.
b) Dookie serum 1/100.
Lane 6 No antibody control.

FIGURE 5: Western blots of SDS extracted antigens from *M. hyopneumoniae* strain Beaufort probed with paired serum and supernatantantibody probes. Fractionation of antigens on SDS Polyacrylamide gel (10.0 %).

Lane 1 a) Pig 453 supernatant.
b) Pig 453 serum 1/100.
Lane 2 a) Pig 105 supernatant.
b) Pig 105 serum 1/100.
Lane 3 a) Pig 1 supernatant.
b) Pig 1 serum 1/100.
Lane 4 a) Pig 15 supernatant.
b) Pig 15 serum 1/100.
Lane 5 a) Dookie supernatant.
b) Dookie serum 1/100.
Lane 6 No antibody control.

FIGURE 6: The entire 48 k gene sequence.

FIGURE 7: tHE 48kDa protein sequence of the 48k gene sequence.

EXAMPLE 1

Mycoplasma hyopneumoniae media

Friss Media

Hovind-Hougen, K., Friss, N.F., Research in Veterinary Science, 1991, 51, pp 155-163, "Morphological & Ultrastructural Studies of M flocculare and M hyopneumoniae in vitro".

250 ml Hanks BSS
140 ml Water
1.5 gm Brain Heart infusion
1.6 gm PPLO Broth w/o CV
Autoclave at 120°C for 20 minutes
18 ml Yeast Extract (100g YSC-2 Sigma in 750 ml)
3.7 ml 0.2% DNA in 0.1% Na_2CL_3
5.14 ml 1% -NAD
0.6 ml 1% Phenol red

Adjust to pH 7.3 to 7.4
Filter through 0.45 um, 0.2 um membrane, store at 4°C.
Add sterile Horse or Pig serum to 20%
and Antibiotics prior to use

Etheridge Media

Etheridge, J.R., Cottew, G.S., Lloyd, L.C., Australian Veterinary Journal, 1979, August 55, pp 356-359, "Isolation of Mycoplasma hyopneumoniae from lesions in experimentally infected pigs".

Materials	For 600 mls
Hanks BSS	18.9 ml
Hartleys Digest broth	1.28 gm
Heart Infusion broth	1.65 gm
Lactalbumin hydrolysate	2.21 gm
Glucose	4.41 gm
Yeast Extract autolysate	8.82 ml
Pig Serum (filtered)	163 ml
1% NAD	6.17 ml
1% Phenol red	1.32 ml
0.2% DNA in 0.1% Na_2CO_3	4.41 ml

Make up to 600 ml with MQ water (about 350 - 400 ml)

Adjust pH to 7.4 and filter through: 3.0 um, 0.8 um, 0.45 um, 0.2 um.

Store at 4°C.

Development of Immune Sows

Cull sows and naive gilt (unmated sow designated Dookie).

Challenged on numerous occasions, with culture grown M. hyopneumoniae and lung homogenate. Given intranasally and intratracheally. Period of challenge - from September, 1991 to 21st January, 1992.

Tiamulin antibiotic given 31st January, 1992 to 4th February, 1992. Rested for approximately 8 weeks.

Infectious Challenge

120 ml of frozen culture of M. hyopneumoniae strain Beaufort, spun down (12,000 xg, 20 min.) and resuspended in 50 ml complete medium and cultured overnight at 37°C. The overnight culture was centrifuged (12,000 xg, 20 min.) and the Mycoplasma cells resuspended in 10 ml serum free Mycoplasma culture medium. The 10 ml of concentrated mycoplasma was administered to anaesthetised immune sows via a catheter to ensure the inoculum was placed into the trachea.

Three of four days post-challenge, the sows were killed, and lymph nod s draining the lungs taken - these included the left and right tracheobronchial lymph nodes, and the lymph nodes located at the bifurcation of the trachea.

Antibody probes were prepared from pig lymph nodes and utilised to detect putative protection antigens as described in Australian Patent Application 49035/90 referred to above. Separate cell cultures were obtained from individual lymph nodes. Culture supernatants were harvested after 5 days of culture.

Antigen Preparation

Mycoplasma hyopneumoniae strain Beaufort was cultured in Etheridge media until the pH had dropped to between 6.8 and 7.0. Cells of M. hyopneumoniae were harvested from culture by centrifugation at 12,000 xg for 20 min., washed 4 times with either sterile PBS or 0.25 M NaCl and then the pelleted cells extracted with one of the following.

(i) Sodium dodecyl sulphate (SDS)

The cell pellet was resuspended in 0.2% SDS and extracted for 2 hours at 37°C. Insoluble material was pelleted from the extract at 12,000 xg for 10 min. and the soluble extract run on SDS-polyacrylamide gel electrophoresis (SDS-PAGE).

(ii) Triton X-114

The method of Bordier (J. Bio. Chem. 1981, 256:1604-1606) was used to selectively extract membrane proteins using the non-ionic detergent Triton X-114.

The cell pellet was resuspended in cold PBS to 2 mg/ml protein and a cold pre-condensed solution of TX-114 added to give a final concentration of 1% (v/v) TX-114. Extraction was achieved by incubation overnight at 4°C with gentle mixing. Insoluble material was removed by centrifugation at 12,000 xg for 20 min. at 4°C. The Triton X-114 soluble membrane proteins were then obtained by achieving a phase separation at 37°C.

Proteins soluble in TX-114 phase were precipitated with 80% ethanol in the presence of carrier dextran (80,000 molecular weight) at -70°C overnight. The proteins were collected by centrifugation at 12,000 xg for 30 min. and dissolved to 500 ug/ml in 4 M urea.

Identification of Antigens

Six antigens were identified utilising the above- mentioned technique. The identified antigens were those that were consistently identified by the antibody probes from the immune cultures and the Dookie gilt. The results are summarised in Table 1.

TABLE 1

Molecular Weight (kD)	Characteristics
110-114	SDS Extracted
90-94	SDS Extracted
72-75	Triton X-114 Extracted
60-64**	SDS Extracted. Partitions to aqueous phase of Triton X-114 extract.
52-54	Triton X-114 Extracted
46-48	Triton X-114 Extracted

** Two antigens of approximate molecular weight 62 kD were identified.

Molecular Weight (kD)	Amino Acid Sequence
46-48	48 K N-Terminal: AGXGQTESGSTSDSKPQAETLKHKV
	48 K CNBR F 1: TIYKPDKVLGKVAVEVLRVLIAKKNKASR
	48 K CNBR F 2: AEQAITKLKLEGFDTQ
	48 K CNBR F 3: KNSQNKIIDLSPEG
52-54	52 K N-Terminal: AGXWAKETTKEEKS
	52 K CNBR F 1: AWVTADGTVN
	52 K CNBR F 2: AIVTADGTVNDNKPNQWVRKY
60-64	62 K N-Terminal: MKLAKLLKGFX (N/L)(M/V) IK
60-64	62 K N-Terminal ADP(F/I)(R/E)Y(V/A)PQG(Q/A)X(M/N)VG

72-75	74 K N-Terminal: AGXLQKNSLLEEVWYLAL
	74 K CNBR F 1: AKNFDFAPSIQGYKKIAHEL
	74 K CNBR F 2: NLKPEQILQLLG
	74 K CNBR F 3: LLKAEXNKXIEEINTXLDN

CNBR - Cyanogen Bromide fragment

X denotes an undetermined amino acid

(A/B) - residue may be A or B

PCR of 48kDa Gene

Polymerase Chain Reaction (PCR) oligonucleotide primers were designed from the amino acid sequences obtained from the N-terminal and internal cyanogen bromide (CNBr) derived peptides. Inosine (I) was substituted at positions of high redundancy. The following primers were used in a standard PCR assay, run on a Bartelt Gene Machine Robotic thermal cycling instrument.

Oligo 48 K CNBr F 1 : ACIAACGACGAGAAGCCICAGGC

T T A A A

Oligo 48 K CNBr F 2 : TTIAGCTTIGTGATIGCCTGCTC

AT A T T

T

Oligo 48 K CNBr F 3 : AGGTCGATGATCTTCCAICC

AA A A T T

T T

The resulting PCR products were visualised on a 1.5% agarose gel, excised, and purified using Prep-a-Gene (BioRad). They were cloned by standard techniques into a dideoxy tailed T-vector (Holton and Graham, Nucleic Acids Research 19: 1156, 1991) and the nucleic acid sequence determined. The PCR product, obtained from the reaction using primers F1 and F2 shown above, was of

approximately 810 base pairs and was shown by sequencing to code for the previously determined amino acid sequence of the purified native 46-48kDa protein.

Genomic clone isolation at 48 k gene

The entire 48k gene and 48kDa protein (Figures 6 and 7) has been isolated and sequenced. The gene was obtained from an *M. hyopneumoniae* genomic library made by digesting genomic DNA with the restriction enzyme CLA I and ligating the fragments into the vector pBluescript (Stratagene). The ligated product was then electroporated into *Escherichia coli* strain SURE (Stratagene) and the cells plated on Luria Broth agar plates containing 100 μg/ml Ampicillin (LB-Amp). The library was screened by DNA hybridisation with a polymerase chain reaction (PCR) product specific for the 48 kDa protein. Positive clones were grown in LB-Amp, the cells harvested and the DNA isolated and partially sequenced for confirmation.

The positive clone pC1-2 was entirely sequenced and the protein sequence deduced. This was compared to the protein sequence obtained from the N terminus and Cyanogen Bromide fragments of the 48 kDa protein to show the that the gene encoded the desired protein.

Adjuvant Selection

Young piglets, 5-7 weeks of age, were immunised with identified antigen(s). The antigens include Triton X-114 extract and identified proteins of 46-48, 52-53, 60-64, 70-75, 90-94 and 110-114 kD, either singly or in combination. An immunising dose of antigen, containing between 5-100 μg protein, was given by intramuscular injection in combination with an adjuvant. An adjuvant is selected from

(i) Seppic Montanide ISA-50

(ii) Quill A and other derivatives of saponin,

(iii) oil in water emulsion employing a mineral oil such as Bayol F/Arlacel A,

(iv) oil in water emulsion employing a vegetable oil such as corn oil,

safflower oil or other with lecithin as emulsifier,

(v) aluminium hydroxide gel, and

(vi) nonionic block polymer such as Pluronic F-127 produced by BASF (U.S.A.).

Immunising doses were given at 2-4 week intervals, the number of doses being dependent on the adjuvant and amount of antigen, but preferably 2 to 3 doses are given.

Adjuvants were treated on the basis of being able to induce antibody titres, as measured by ELISA, and by assessment of induced cell-mediated immunity as tested by Delayed-Type Hypersensitivity (DTH) reaction.

The results clearly show that mineral-oil type adjuvants are consistently superior at inducing antibody titres and DTH responses (Table 2). In particular an adjuvant marketed under trade designation Montanide ISA-50 and available from Seppic, Paris, France has been found to be suitable.

TABLE 2

GROUP	Animal Number	DTH 24 Hour Response	DTH 48 Hour Response	Antibody Levels (450 nm)
CONTROL (Unvaccinated)	19	0	0	0.061
	11	0	0	0.010
	1	-	-	0.005
	15	0	0	0.038
	7	0	0	0.005
QUIL A	18	+	0	0.753
	25	+	0	0.788
	17	0	0	0.638
	168	_	±	0.642
VEG. OIL	169	+++	0	0.316
	22	0	0	0.621
	4	+	0	0.666
	6	+	+	0.239
	13	+++	++	0.457
MIN. OIL	14	+++	++	1.086
	5	+++	++	1.024
	23	+++	+	0.864
	16	+++	0	0.975
	21	+	±	0.954

TABLE 2: Antibody levels and DTH responses in pigs measured 2 weeks after the third injection of antigen from M. hyopneumoniae. (- = no response; ± = faint reddening; + = faint reddening and swelling; ++ = reddening; +++ = swelling with or without reddening).

Protection Pen Trial

Groups of 9 young piglets, 6 weeks of age, were immunised with purified and semi-purified antigens as shown in Table 3 below. The antigens were purified on reversed-phase HPLC using a formic acid solvent system with an acetonitrile gradient.

Antigens were resolubilised in 4 Molar urea before incorporation in mineral oil adjuvant.

The immunisation schedule is as shown in Table 2.

TABLE 3

Protocol for Protection Trial of Antigens of Mycoplasma Hyopneumonia

VACCINATIONS & BLEEDS

Treatment	Day Number
1st Vaccination	0
2nd Vaccination	14
3rd Vaccination	50
Infectious Challenge	64
Slaughter	91

ANTIGEN DOSES

Partly Purified 62 kD	1st & 2nd Vaccns. 50μg COMPLEX ANTIGEN/DOSE 3rd Vaccn. - 220μg PARTIALLY PURIFIED ANTIGEN/DOSE
(Purified)74+52kD	1st Vaccn. 20μg total protein/DOSE 2nd Vaccn. 13μg total protein/DOSE 3rd Vaccn. 17μg total protein/DOSE
(Purified) 48KD	1st Vaccn. 20μg/DOSE 2nd Vaccn. 18μg/DOSE 3rd Vaccn. 27μg/DOSE

ALL PROTEIN ESTIMATIONS DONE BY "BCA" PROTEIN ASSAY (Pierce, Illinois, U.S.A.

Protection from infection with Mycoplasma hyopneumoniae was assessed by infectious challenge 2 weeks after the final immunisation. Infectious challenge was achieved by intranasal administration of 10ml of a 10% (w/v) lung homogenate, prepared from infected lung, and by housing test piglets with

previously infected piglets. Four weeks after infectious challenge, the animals were killed and the extent and degree of lung lesions assessed (Table 4).

TABLE 4

Pen Trial of Antigens of Mycoplasma Hyopneumoniae

Group No.	No. Pneumonia Free (%)	Median Lung Lesion Score	% Reduction (from Median)
Controls	1 (11)	13	0%
62 kD	0 (0)	5	61%
74+52 kD	3 (33)	6.75	48%
48 kD	2 (22)	6.25	52%

REFERENCE

Warren H.S. and Chedid, L.A., Future Prospects for Vaccine Adjuvants CRC Critical Reviews in Immunology 8 : 83-108, 1988.

Finally, it is to be understood that various other modifications and/or alterations may be made without departing from the spirit of the present invention as outlined herein.

CLAIMS:

1. A putative protective antigen against a Mycoplasma, prepared by a method including

providing

a sample of a Mycoplasma;

an antibody probe including at least one antibody against a Mycoplasma produced by a method including;

providing a biological sample taken a short time after an immune animal has been challenged with a Mycoplasma or Mycoplasma extract taken from the infection site or an area of a lesion or an area close to the infection site or lesion;

isolating cells from the biological sample;

culturing cells in vitro in a suitable culture medium; and

harvesting antibodies produced from said cells;

probing the Mycoplasma sample with the antibody probe to detect at least one antigen; and

isolating the antigen detected.

2. A putative protective antigen according to claim 1 wherein the Mycoplasma is Mycoplasma hyopneumoniae.

3. A putative protective antigen against Mycoplasma hyopneumoniae, or related infections, selected from the group of antigens having approximate molecular weights of 110-114, 90-94, 72-75, 60-64, 52-54 and 46-48 kilodaltons (kD), as herein described, mutants, derivatives and fragments thereof.

4. A putative protective antigen according to claim 3 which is a surface protein.

5. A putative protective antigen according to claim 3 or 4 which is a surface lipo-protein or membrane protein.

6. A putative protective antigen according to any one of claims 3-5 having approximate molecular weight of 110-114, 90-94, 74, 62, 52 and 48 kD.

7. A putative protective antigen according to claim 3 wherein the antigen in the 72-75 kD region contains the following N-terminal amino acid sequence:

AGXLQKNSLLEEVWYLAL

8. A putative protective antigen according to claim 7 further including one or more of the following N-terminal amino acid sequences:

AKNFDFAPSIQGYKKIAHEL

NLKPEQILQLLG

LLKAEXNKXIEEINTXLDN

9. A putative protective antigen according to claim 3 wherein the antigen in the 60-64 kD region contains the following N-terminal amino acid sequence:

MKLAKLLKGFX(N/L)(M/V)IK

ADP(F/I)(R/E)Y(V/A)PQG(Q/A)X(M/N)VG

10. A putative protective antigen according to claim 3 wherein the antigen in the 52-54 kD region contains the following N-terminal amino acid sequence:

AGXWAKETTKEEKS

11. A putative protective antigen according to claim 10 further including one or more of the following N-terminal amino sequences:

AWVTADGTVN

AIVTADGTVNDNKPNQWVRKY.

12. A putative protective antigen according to claim 3 wherein the antigen in the 46-48 kD region contains the following N-terminal amino acid sequence:

AGXGQTESGSTSDSKPQAETLKHKV

13. A putative protective antigen according to claim 12 further including one or more of the following internal amino acid sequences:

TIYKPDKVLGKVAVEVLRVLIAKKNKASR

AEQAITKLKLEGFDTQ

KNSQNKIIDLSPEG

14. An isolated nucleic acid fragment encoding a putative protective antigen against Mycoplasma hyopneumoniae or related infections, said nucleic acid fragment including the following sequence, mutants, derivatives, recombinants and fragments thereof:

10 1234567890	20 1234567890	30 1234567890	40 1234567890	50 1234567890	
ATGAAAAAAA	TGCCACTATA	CCAGAGGAAA	GAGCAGTATA	TAAAATAATT	50
AAAATTACAT	TTTCTTCATT	TGCGCCAGAA	TTTTTAAGAA	TTAGTACATT	100
AAAAAGTAGA	ACAAAAGTTA	TTAATGTAAA	CATTAGCGCA	ATCCTTAAGA	150
AAAAATTAAA	AGTTTTATCT	ATTTTTTTTA	ATCGAAATCC	AACCAGGCAT	200
AAATCTTTGT	CAGTATTTAT	CAAGTCGGTA	TTTTTTCATT	ATTTCTACTA	250
AAATATTATT	TGAATTTGCA	TTTTCCATAA	TCTAAAATTT	TACATTTTTT	300
TATAACAATT	TTTAAAAATT	ACTCTTTAAT	TTATAGTATT	TTTTTATTTT	350
TTAGTCTAAA	TTATAAAATT	ATCTTGAATT	TTATTTGAAT	TTTTATAATT	400
TAGTACTAAA	AAATACAAAT	ATTTTTTCCT	ATTCTAAGAA	AAATTCATTT	450
TTTAAAAAAA	ATTGATTTTT	ATAGTATAAT	TTGTTTGTAT	AATTGAATTA	500
ACTTGATTTG	AAAGGGAACA	AAATGAAAAA	AATGCTTAGA	AAAAAATTCT	550
TGTATTCATC	AGCTATTTAT	GCAACTTCGC	TTGCATCAAT	TATTGCATTT	600
GTTGCAGCAG	GTTGTGGACA	GACAGAATCA	GGTTCAACTT	CTGATTCTAA	650
ACCACAAGCC	GAGACGCTAA	AACATAAAGT	AAGTAATGAT	TCTATTCGAA	700
TAGCACTAAC	CGATCCGGAT	AATCCTCGAT	GAATTAGTGC	CCAAAAAGAT	750
ATTATTTCTT	ATGTTGATGA	AACAGAGGCA	GCAACTTCAA	CAATTACAAA	800
AAACCAGGAT	GCACAAAATA	ACTGACTCAC	TCAGCAAGCT	AATTTAAGCC	850
CAGCGCCAAA	AGGATTTATT	ATTGCCCCTG	AAAATGGAAG	TGGAGTTGGA	900
ACTGCTGTTA	ATACAATTGC	TGATAAAGGA	ATTCCGATTG	TTGCCTATGA	950
TCGACTAATT	ACTGGATCTG	ATAAATATGA	TTGGTATGTT	TCTTTTGATA	1000
ATGAAAAAGT	TGGTGAATTA	CAAGGTCTTT	CACTTGCTGC	GGGTCTATTA	1050
GGAAAAGAAG	ATGGTGCTTT	TGATTCAATT	GATCAAATGA	ATGAATATCT	1100
AAAATCACAT	ATGCCCCAAG	AGACAATTTC	TTTTTATACA	ATCGCGGGTT	1150
CCCAAGATGA	TAATAATTCC	CAATATTTTT	ATAATGGTGC	AATGAAAGTA	1200
CTTAAAGAAT	TAATGAAAAA	TTCGCAAAAT	AAAATAATTG	ATTTATCTCC	1250
TGAAGGCGAA	AATGCTGTTT	ATGTCCCAGG	ATGAAATTAT	GGAACTGCCG	1300
GTCAAAGAAT	CCAATCTTTT	CTAACAATTA	ACAAAGATCC	AGCAGGTGGT	1350
AATAAAATCA	AAGCTGTTGG	TTCAAAACCA	GCTTCTATTT	TCAAAGGATT	1400
TCTTGCCCCA	AATGATGGAA	TGGCCGAACA	AGCAATCACC	AAATTAAAAC	1450
TTGAAGGGTT	TGATACCCAA	AAAATCTTTG	TAACTCGTCA	AGATTATAAT	1500
GATAAAGCCA	AAACTTTTAT	CAAAGACGGC	GATCAAAATA	TGACAATTTA	1550

TAAACCTGAT	AAAGTTTTAG	GAAAAGTTGC	TGTTGAAGTT	CTTCGGGTTT	1600
TAATTGCAAA	GAAAAATAAA	GCATCTAGAT	CAGAAGTCGA	AAACGAACTA	1650
AAAGCAAAAC	TACCAAATAT	TTCATTTAAA	TATGATAATC	AAACATATAA	1700
AGTACAAGGT	AAAAATATTA	ATACAATTTT	AGTAAGTCCA	GTAATTGTTA	1750
CAAAAGCTAA	TGTTGATAAT	CCTGATGCCT	AA		1782

15. An isolated nucleic acid fragment according to claim 14 encoding a putative protective antigen wherein the antigen is in the 46-48 kD region including the following nucleic acid sequence, mutants, derivatives, recombinants and fragments thereof:

10 1234567890	20 1234567890	30 1234567890	40 1234567890	50 1234567890	
ATGAAAAAAA	TGCCACTATA	CCAGAGGAAA	GAGCAGTATA	TAAAATAATT	50
AAAATTACAT	TTTCTTCATT	TGCGCCAGAA	TTTTTAAGAA	TTAGTACATT	100
AAAAAGTAGA	ACAAAAGTTA	TTAATGTAAA	CATTAGCGCA	ATCCTTAAGA	150
AAAAATTAAA	AGTTTTATCT	ATTTTTTTTA	ATCGAAATCC	AACCAGGCAT	200
AAATCTTTGT	CAGTATTTAT	CAAGTCGGTA	TTTTTTCATT	ATTTCTACTA	250
AAATATTATT	TGAATTTGCA	TTTTCCATAA	TCTAAAATTT	TACATTTTTT	300
TATAACAATT	TTTAAAAATT	ACTCTTTAAT	TTATAGTATT	TTTTTATTTT	350
TTAGTCTAAA	TTATAAAATT	ATCTTGAATT	TTATTTGAAT	TTTTATAATT	400
TAGTACTAAA	AAATACAAAT	ATTTTTTCCT	ATTCTAAGAA	AAATTCATTT	450
TTTAAAAAAA	ATTGATTTTT	ATAGTATAAT	TTGTTTGTAT	AATTGAATTA	500
ACTTGATTTG	AAAGGGAACA	AAATGAAAAA	AATGCTTAGA	AAAAAATTCT	550
TGTATTCATC	AGCTATTTAT	GCAACTTCGC	TTGCATCAAT	TATTGCATTT	600
GTTGCAGCAG	GTTGTGGACA	GACAGAATCA	GGTTCAACTT	CTGATTCTAA	650
ACCACAAGCC	GAGACGCTAA	AACATAAAGT	AAGTAATGAT	TCTATTCGAA	700
TAGCACTAAC	CGATCCGGAT	AATCCTCGAT	GAATTAGTGC	CCAAAAAGAT	750
ATTATTTCTT	ATGTTGATGA	AACAGAGGCA	GCAACTTCAA	CAATTACAAA	800
AAACCAGGAT	GCACAAAATA	ACTGACTCAC	TCAGCAAGCT	AATTTAAGCC	850
CAGCGCCAAA	AGGATTTATT	ATTGCCCCTG	AAAATGGAAG	TGGAGTTGGA	900
ACTGCTGTTA	ATACAATTGC	TGATAAAGGA	ATTCCGATTG	TTGCCTATGA	950
TCGACTAATT	ACTGGATCTG	ATAAATATGA	TTGGTATGTT	TCTTTTGATA	1000
ATGAAAAAGT	TGGTGAATTA	CAAGGTCTTT	CACTTGCTGC	GGGTCTATTA	1050
GGAAAAGAAG	ATGGTGCTTT	TGATTCAATT	GATCAAATGA	ATGAATATCT	1100
AAAATCACAT	ATGCCCCAAG	AGACAATTTC	TTTTTATACA	ATCGCGGGTT	1150
CCCAAGATGA	TAATAATTCC	CAATATTTTT	ATAATGGTGC	AATGAAAGTA	1200
CTTAAAGAAT	TAATGAAAAA	TTCGCAAAAT	AAAATAATTG	ATTTATCTCC	1250
TGAAGGCGAA	AATGCTGTTT	ATGTCCCAGG	ATGAAATTAT	GGAACTGCCG	1300
GTCAAAGAAT	CCAATCTTTT	CTAACAATTA	ACAAAGATCC	AGCAGGTGGT	1350
AATAAAATCA	AAGCTGTTGG	TTCAAAACCA	GCTTCTATTT	TCAAAGGATT	1400
TCTTGCCCCA	AATGATGGAA	TGGCCGAACA	AGCAATCACC	AAATTAAAAC	1450
TTGAAGGGTT	TGATACCCAA	AAAATCTTTG	TAACTCGTCA	AGATTATAAT	1500
GATAAAGCCA	AAACTTTTAT	CAAAGACGGC	GATCAAAATA	TGACAATTTA	1550
TAAACCTGAT	AAAGTTTTAG	GAAAAGTTGC	TGTTGAAGTT	CTTCGGGTTT	1600
TAATTGCAAA	GAAAAATAAA	GCATCTAGAT	CAGAAGTCGA	AAACGAACTA	1650
AAAGCAAAAC	TACCAAATAT	TTCATTTAAA	TATGATAATC	AAACATATAA	1700
AGTACAAGGT	AAAAATATTA	ATACAATTTT	AGTAAGTCCA	GTAATTGTTA	1750
CAAAAGCTAA	TGTTGATAAT	CCTGATGCCT	AA		1782

16. A method for producing an antibody against a Mycoplasma including

providing a biological sample taken a short time after an immune animal has been challenged with a Mycoplasma or Mycoplasma extract taken from the infection site or an area of a lesion or an area close to the infection site or lesion;

isolating cells from the biological sample;

culturing cells in vitro in a suitable culture medium; and

harvesting antibodies produced from said cells.

17. A method according to claim 16 wherein the biological sample is taken at a predetermined time after the animal has been challenged with a Mycoplasma, preferably 2 to 7 days after challenge.

18. A method according to claim 16 wherein the culturing of cells in vitro further includes addition of helper factors to the culture, said helper factors selected from the group including cytokines used alone or in combination, including Interleukin 1, 2, 3, 4, 5, 6, 7 and 8, colony stimulating factors, interferons and any other factors that may be shown to have an enhancing effect on specific B cell secretion.

19. A method according to any one of claims 16-18 further including a cell activation step including activating the cells isolated to proliferate and secrete and/or release antibodies

said cell activation step including adding a cell activating agent to the culture medium, said cell activating agent selected from the group including mitogens as herein described and helper factors produced by leukocytes, or their synthetic equivalents or combinations thereof.

20. A method according to any one of claims 16-19 wherein the antibody is in the form of the supernatant harvested from the culture medium.

21. An antibody against a Mycoplasma prepared according to the method of any one of claims 16-20.

22. A method of identifying a putative protective antigen associated with a Mycoplasma, preferably Mycoplasma hyopneumoniae, said method including providing

a sample of a Mycoplasma; and

an antibody probe including at least one antibody against a Mycoplasma;

probing the Mycoplasma sample with the antibody probe to detect at least one antigen; and

isolating the antigen detected.

23. A method of purifying a putative protective antigen associated with a Mycoplasma, preferably Mycoplasma hyopneumoniae, said method including providing

a crude antigen mixture; and

an antibody against a Mycoplasma immobilized on a suitable support;

subjecting the crude antigen mixture to affinity chromatography utilizing the immobilized antibody; and

isolating the purified antigen so formed.

24. A method for preparing a synthetic antigenic polypeptide against Mycoplasma, preferably Mycoplasma hyopneumoniae, which method includes providing

a cDNA library or genomic library derived from a sample of Mycoplasma; and

an antibody probe including an antibody prepared according to claim 16;

generating synthetic polypeptides from the cDNA library or genomic library;

probing the synthetic polypeptides with the antibody probe; and

isolating the synthetic antigenic polypeptide detected thereby.

25. A method according to claim 24 wherein th antibody probe includes an antibody raised against an antigen against Mycoplasma hyopneumoniae, or related infections, selected from the group of antigens having approximate molecular weights of 110-114, 90-94, 72-75, 60-64, 52-54 and 46-48 kilodaltons (kD), as herein described, mutants, derivatives and fragments thereof.

26. A synthetic putative protective antigen in the 72-75 kD region produced by a method according to claim 24 or 25 having an N-terminal amino acid sequence:

AGXLQKNSLLEEVWYLAL

27. A synthetic putative protective antigen according to claim 26 further including internal amino acid sequences:

AKNFDFAPSIQGYKKIAHEL

NLKPEQILQLLG

LLKAEXNKXIEEINTXLDN

28. A synthetic putative protective antigen in the 60-64 kD region produced by a method according to claim 24 or 25 having an N-terminal amino acid sequence:

MKLAKLLKGFX(N/L)(M/V)IK

ADP(F/I)(R/E)Y(V/A)PQG(Q/A)X(M/N)VG

29. A synthetic putative protective antigen in the 52-54 kD region produced by a method according to claim 24 or 25 having an n-terminal amino acid sequence;

AGXWAKETTKEEKS

30. A synthetic putative protective antigen according to claim 29 further including internal amino acid sequences:

AWVTADGTVN

AIVTADGTVNDNKPNQWVRKY.

31. A synthetic putative protective antigen in the 46-48 kD region produced by a method according to claim 24 or 25 having an N-terminal amino acid sequence:

AGXGQTESGSTSDSKPQAETLKHKV

32. A synthetic putative protective antigen according to claim 31 further including internal amino acid sequences:

TIYKPDKVLGKVAVEVLRVLIAKKNKASR

AEQAITKLKLEGFDTQ

KNSQNKIIDLSPEG

33. A vaccine or veterinary composition including a prophylactically effective amount of at least one putative protective antigen against a Mycoplasma according to any one of claims 1-13.

34. A vaccine or veterinary composition according to claim 33 including a plurality of putative protective antigens selected from antigens having approximate molecular weights of 110-114, 90-94, 72-75, 60-64, 52-54 and 46-48 kilodaltons.

35. A vaccine or veterinary composition including an antibody against a Mycoplasma according to claim 21.

36. A diagnostic kit including a diagnostic antigen or fragment thereof according to any one of claims 1-13 and 26-32.

37. A method for preventing or treating a Mycoplasma infection, which method including administering to an animal a prophylactically or therapeutically effective amount of at least one putative protective antigen according to any one of claims 1-13.

38. An isolated DNA fragment encoding a putative protective antigen against Mycoplasma or related infections, said DNA fragment having a nucleic acid sequence according to Figure 6 or an homologous sequence, and functionally active fragments, mutant, variant or recombinant thereof.

39. A clone including a DNA fragment according to claim 38.

40. A clone according to claim 39 which is clone pC1-2 as hereinbefore described.

41. An amino acid sequence or functional equivalent thereof encoded by the DNA fragment according to claim 38.

42. An amino acid sequence or functional equivalent thereof having the amino acid sequence of Figure 7.

43. A putative protective antigen or antibody substantially as hereinbefore described with reference to the examples.

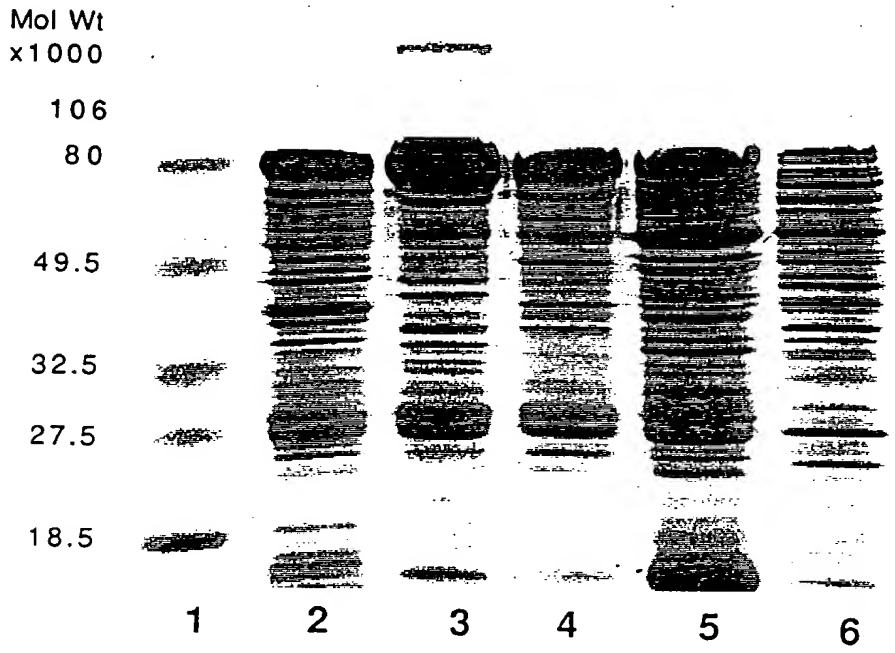
Mol Wt
x1000
106
80
49.5
32.5
27.5
18.5
1
2
3
4
5
6

FIG. 1

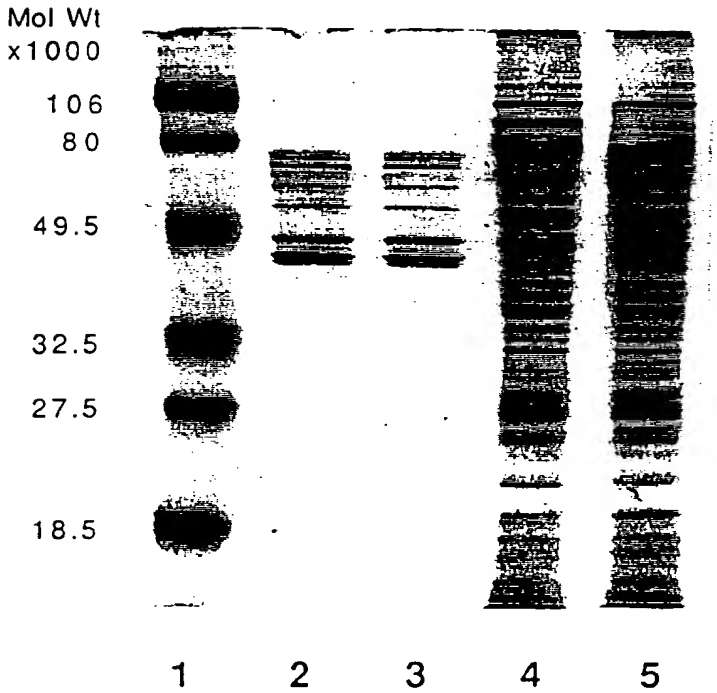
Mol Wt
x1000
106
80
49.5
32.5
27.5
18.5
1
2
3
4
5

FIG. 2

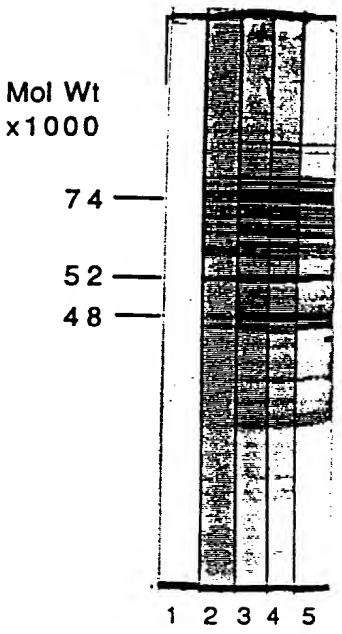

FIG. 3

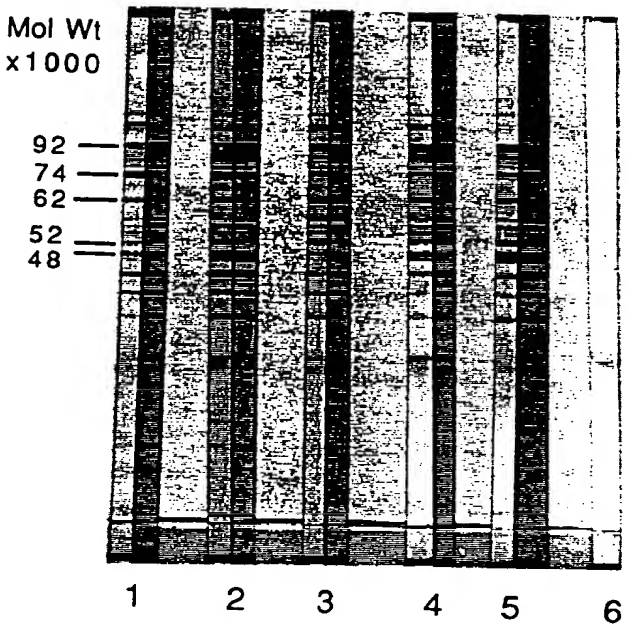

FIG. 4

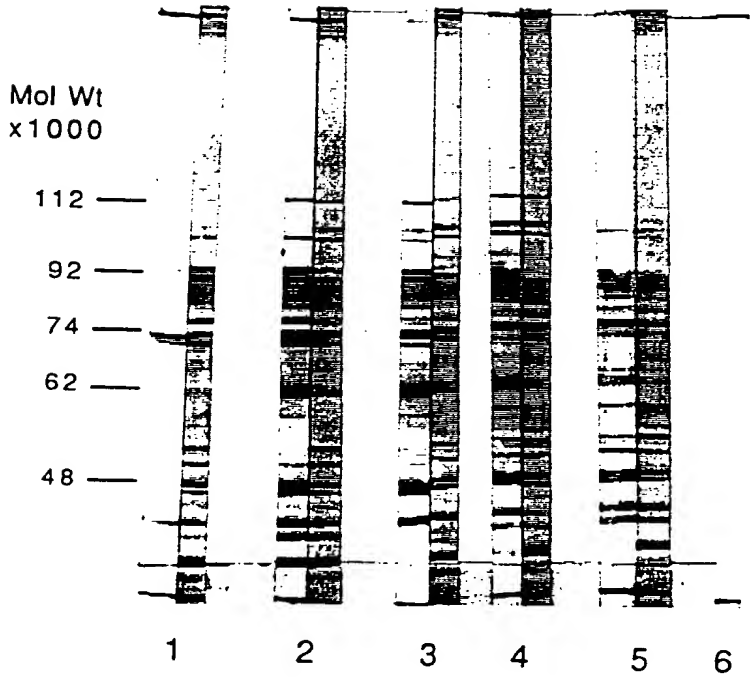

FIG. 5

10	20	30	40	50	
1234567890	1234567890	1234567890	1234567890	1234567890	
ATGAAAAAAA	TGCCACTATA	CCAGAGGAAA	GAGCAGTATA	TAAAATAATT	50
AAAATTACAT	TTTCTTCATT	TGCGCCAGAA	TTTTTAAGAA	TTAGTACATT	100
AAAAAGTAGA	ACAAAAGTTA	TTAATGTAAA	CATTAGCGCA	ATCCTTAAGA	150
AAAAATTAAA	AGTTTTATCT	ATTTTTTTTA	ATCGAAATCC	AACCAGGCAT	200
AAATCTTTGT	CAGTATTTAT	CAAGTCGGTA	TTTTTTCATT	ATTTCTACTA	250
AAATATTATT	TGAATTTGCA	TTTTCCATAA	TCTAAAATTT	TACATTTTTT	300
TATAACAATT	TTTAAAAATT	ACTCTTTAAT	TTATAGTATT	TTTTTATTTT	350
TTAGTCTAAA	TTATAAAATT	ATCTTGAATT	TTATTTGAAT	TTTTATAATT	400
TAGTACTAAA	AAATACAAAT	ATTTTTTCCT	ATTCTAAGAA	AAATTCATTT	450
TTTAAAAAAA	ATTGATTTTT	ATAGTATAAT	TTGTTTGTAT	AATTGAATTA	500
ACTTGATTTG	AAAGGGAACA	AAATGAAAAA	AATGCTTAGA	AAAAAATTCT	550
TGTATTCATC	AGCTATTTAT	GCAACTTCGC	TTGCATCAAT	TATTGCATTT	600
GTTGCAGCAG	GTTGTGGACA	GACAGAATCA	GGTTCAACTT	CTGATTCTAA	650
ACCACAAGCC	GAGACGCTAA	AACATAAAGT	AAGTAATGAT	TCTATTCGAA	700
TAGCACTAAC	CGATCCGGAT	AATCCTCGAT	GAATTAGTGC	CCAAAAAGAT	750
ATTATTTCTT	ATGTTGATGA	AACAGAGGCA	GCAACTTCAA	CAATTACAAA	800
AAACCAGGAT	GCACAAAATA	ACTGACTCAC	TCAGCAAGCT	AATTTAAGCC	850
CAGCGCCAAA	AGGATTTATT	ATTGCCCCTG	AAAATGGAAG	TGGAGTTGGA	900
ACTGCTGTTA	ATACAATTGC	TGATAAAGGA	ATTCCGATTG	TTGCCTATGA	950
TCGACTAATT	ACTGGATCTG	ATAAATATGA	TTGGTATGTT	TCTTTTGATA	1000
ATGAAAAAGT	TGGTGAATTA	CAAGGTCTTT	CACTTGCTGC	GGGTCTATTA	1050
GGAAAAGAAG	ATGGTGCTTT	TGATTCAATT	GATCAAATGA	ATGAATATCT	1100
AAAATCACAT	ATGCCCCAAG	AGACAATTTC	TTTTTATACA	ATCGCGGGTT	1150
CCCAAGATGA	TAATAATTCC	CAATATTTTT	ATAATGGTGC	AATGAAAGTA	1200
CTTAAAGAAT	TAATGAAAAA	TTCGCAAAAT	AAAATAATTG	ATTTATCTCC	1250
TGAAGGCGAA	AATGCTGTTT	ATGTCCCAGG	ATGAAATTAT	GGAACTGCCG	1300
GTCAAAGAAT	CCAATCTTTT	CTAACAATTA	ACAAAGATCC	AGCAGGTGGT	1350
AATAAAATCA	AAGCTGTTGG	TTCAAAACCA	GCTTCTATTT	TCAAAGGATT	1400
TCTTGCCCCA	AATGATGGAA	TGGCCGAACA	AGCAATCACC	AAATTAAAAC	1450
TTGAAGGGTT	TGATACCCAA	AAAATCTTTG	TAACTCGTCA	AGATTATAAT	1500
GATAAAGCCA	AAACTTTTAT	CAAAGACGGC	GATCAAAATA	TGACAATTTA	1550
TAAACCTGAT	AAAGTTTTAG	GAAAAGTTGC	TGTTGAAGTT	CTTCGGGTTT	1600
TAATTGCAAA	GAAAAATAAA	GCATCTAGAT	CAGAAGTCGA	AAACGAACTA	1650
AAAGCAAAAC	TACCAAATAT	TTCATTTAAA	TATGATAATC	AAACATATAA	1700
AGTACAAGGT	AAAAATATTA	ATACAATTTT	AGTAAGTCCA	GTAATTGTTA	1750
CAAAAGCTAA	TGTTGATAAT	CCTGATGCCT	AA		1782

FIG. 6

```
          10         20         30         40         50
1234567890 1234567890 1234567890 1234567890 1234567890

MKKMLRKKFL YSSAIYATSL ASIIAFVAAG CGQTESGSTS DSKPQAETLK      50
HKVSNDSIRI ALTDPDNPRW ISAQKDIISY VDETEAATST ITKNQDAQNN     100
WLTQQANLSP APKGFIIAPE NGSGVGTAVN TIADKGIPIV AYDRLITGSD     150
KYDWYVSFDN EKVGELQGLS LAAGLLGKED GAFDSIDQMN EYLKSHMPQE     200
TISFYTIAGS QDDNNSQYFY NGAMKVLKEL MKNSQNKIID LSPEGENAVY     250
VPGWNYGTAG QRIQSFLTIN KDPAGGNKIK AVGSKPASIF KGFLAPNDGM     300
AEQAITKLKL EGFDTQKIFV TRQDYNDKAK TFIKDGDQNM TIYKPDKVLG     350
KVAVEVLRVL IAKKNKASRS EVENELKAKL PNISFKYDNQ TYKVQGKNIN     400
TILVSPVIVT KANVDNPDA                                       419
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FIG. 7

INTERNATIONAL SEARCH REPORT

Internati nal Application N .
PCT/AU 96/00149

A. CLASSIFICATION OF SUBJECT MATTER

Int Cl[6]: C07K 16/12; C12N 15/31; C12P 21/00, 21/02, 21/08; G01N 33/53, 33/531; A61K 39/04; C07K 14/30

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)
IPC As Above

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched
AU: IPC As Above

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)
WPAT, BIOT, JAPIO: MYCOPLASM: AND ANTIGEN#
CASM: MULTI-SEQUENCES

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	AU,A, 70685/87 (CETUS CORP) 1 October 1987 Example 1, Claims	1,16,17,20-23,33 35-37,43
X	AU,B, 49035/90 (640364) (UNIVERSITY OF MELBOURNE et al) 11 October 1990 Claims	1,2,16,20,22
X	AU,A, 76820/91 (SYNERGEN, INC.) 17 October 1991 Fig. 1, 4, 6 and 7, Claim 3	14,38,39,41,42

[X] Further documents are listed in the continuation of Box C

[X] See patent family annex

* Special categories of cited documents:

"A" document defining the general state of the art which is not considered to be of particular relevance

"E" earlier document but published on or after the international filing date

"L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)

"O" document referring to an oral disclosure, use, exhibition or other means

"P" document published prior to the international filing date but later than the priority date claimed

"T" later document published after the international filing date r priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention

"X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone

"Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art

"&" document member of the same patent family

Date f the actual completion of the international search	Date of mailing of the international search report
6 May 1996	15.05.96
Name and mailing address of the ISA/AU AUSTRALIAN INDUSTRIAL PROPERTY ORGANISATION PO BOX 200 WODEN ACT 2606 AUSTRALIA Facsimile No.: (06) 285 3929	Authorized officer **BARRY SPENCER** Telephone No.: (06) 283 2284

INTERNATIONAL SEARCH REPORT

International Application No.
PCT/AU 96/00149

C (Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
PX	AU,A, 17602/95 (SYNERGEN, INC.) 26 October 1995 Fig. 1, 4, 6 and 7 P 13 L14-P14 L5	14,38,39,41,42
X	AU,B, 49599/90 (638970) (AUSPHARM INTERNATIONAL LTD.) 26 July 1990 Examples 4 and 5	16,17,20,21,35
X	US,A, 4894332 (SCHALLER et al) 16 January 1990 Examples	1,2,14,16,17, 20-22,24,33, 35,37-39,41,42
X	US,A, 5252328 (FAULDS et al) 12 October 1993 Examples 1 to 4	1-17,20-23, 33-35,37
X	US,A, 5240706 (FAULDS) 31 August 1993 Examples 1 and 2, Claims	1-6,14-16,21 22,24,25,33-35 37,38,41,42
X	EP,A, 0475185 (NIPPON FLOUR MILLS CO., LTD.) Refer Example 1, Claims	1-6,12-17, 20-23,33-35, 37-39,41,42
PX	Journal of Bacteriology, Vol. 177, No. 7, April 1995, pp1915-1917, "Molecular Cloning of a 46-Kilodalton Surface Antigen (P46) Gene from Mycoplasma hyopneumoniae: Direct evidence of CGG Codon Usage for Arginine", Futo et al	3-6,12-15,24 25,31,32,38, 39,41,42
PX	Journal of Clinical Microbiology, Vol. 33, No. 3, March 1995, pp 680-683, "Recombinant 46-Kilodalton Surface Antigen (P46) of Mycoplasma hyopneumoniae Expressed in Escherichia Coli Can Be Used for early Specific Diagnosis of Mycoplasmal Pneumonia of Swine by Enzyme-Linked Immunosorbent assay", Futo et al	3-6, 12-15,38,39, 41,42
X	Infection and Immunity. Vol. 49, No. 2, pp329-335, "surface Proteins of Mycoplasma hyopneumoniae Identified from an Esherichia coli Expression Plasmid Library", Klinkert et al	3-6,12-15,38 39,41,42
A	EP,A, 0571648 (WENG) 1 December 1993	
A	Derwent abstract Accession No. 88-010509, Class 503, JP,A, 62-273455 (NORIINSHO KK) 27 November 1987	
A	Derwent Abstract Accession No. 90-241949, Class 503, JP,A, 02-167079 (NIPPON SEIFUN KK) 27 June 1990	
A	Derwent abstract Accession No. 95-203749, Class B04, C06, D16, JP,A, 07-118167 (ZENKOKU NOGYO KYODO KUMIAI RENGOKAI) 9 May 1995	

INTERNATIONAL SEARCH REPORT

International Application No.
PCT/AU 96/00149

This Annex lists the known "A" publication level patent family members relating to the patent documents cited in the above-mentioned international search report. The Australian Patent Office is in no way liable for these particulars which are merely given for the purpose of information.

Patent Document Cited in Search Report		Patent Family Member					
AU	87/70685	EP	254384	JP	63000298		
AU	90/49035	AT	134386	BR	9000451	CA	2008808
		CN	1046188	DE	69025414	EP	381427
		FI	900498	IL	93234	JP	3087199
		NO	178893	NZ	232279	ZA	9000766
AU	91/76820	AU	17602/95	CA	2078131	EP	527771
		FI	924428	HU	65827	JP	5506984
		NO	923828	WO	9115593	US	5459048
		AT	135048	AU	622855	DE	3751727
		DK	1608/88	EP	315637	HU	208550
		IL	83324	JP	1503735	WO	8800977
AU	95/17602	AU	76820/91	CA	2078131	EP	527771
		FI	924428	HU	65827	JP	5506984
		NO	923828	WO	9115593	US	5459048
		AT	135048	AU	622855	DE	3751727
		DK	1608/88	EP	315637	HU	208550
		IL	83324	JP	1503753	WO	8800977
AU	90/49599	EP	454735	NZ	232190	WO	9007935
		ZA	9000474				
US	4894332	CA	1301677	CN	86102858	EP	196215
		JP	61274687				

END OF ANNEX

This Annex lists the known "A" publication level patent family members relating to the patent documents cited in the above-mentioned international search report. The Australian Patent Office is in no way liable for these particulars which are merely given for the purpose of information.

Patent Document Cited in Search Report		Patent Family Member					
US	5252328	CA	1321142	CN	88101554	DK	1674/88
		EP	283840	HU	203672	IE	61626
		JP	63258427	PT	87041		
US	5240706	AT	134705	DE	68925769	EP	359919
		JP	2291271				
EP	475185	JP	5091882				

END OF ANNEX

PATENT COOPERATION TREATY

PCT

INTERNATIONAL PRELIMINARY EXAMINATION REPORT

(PCT Article 36 and Rule 70)

Applicant's or agent's file reference IRN 444950	**FOR FURTHER ACTION**	See Notification of Transmittal of International Preliminary Examination Report (Form PCT/IPEA/416).
International application No. **PCT/AU 96/00149**	International filing date 15 March 1996	Priority Date 16 March 1995
International Patent Classification (IPC) or national classification and IPC **Int. Cl.**[6] C07K 16/12, 14/30; C12N 15/31; C12P 21/00, 21/02, 21/08; G01N 33/53, 33/531; A61K 39/04		
Applicant (1) UNIVERSITY OF MELBOURNE (2) WALKER, John; LEE, Rogan; DOUGHTY, Stephen William		

1. This international preliminary examination report has been prepared by this International Preliminary Examining Authority and is transmitted to the applicant according to Article 36.

2. This REPORT consists of a total of **6** sheets, including this cover sheet.

☐ This report is also accompanied by ANNEXES, i.e., sheets of the description, claims and/or drawings which have been amended and are the basis for this report and/or sheets containing rectifications made before this Authority (see Rule 70.16 and Section 607 of the Administrative Instructions under the PCT).

These annexes consist of a total of sheet(s).

3. This report contains indications relating to the following items:

I	☒	Basis of the report
II	☐	Priority
III	☐	Non-establishment of opinion with regard to novelty, inventive step and industrial applicability
IV	☒	Lack of unity of invention
V	☒	Reasoned statement under Article 35(2) with regard to novelty, inventive step or industrial applicability; citations and explanations supporting such statement
VI	☒	Certain documents cited
VII	☐	Certain defects in the international application
VIII	☒	Certain observations on the international application

Date of submission of the demand 4 September 1996	Date of completion of the report 23 December 1996
Name and mailing address of the IPEA/AU AUSTRALIAN INDUSTRIAL PROPERTY ORGANISATION PO BOX 200 WODEN ACT 2606 AUSTRALIA Facsimile No. (06) 285 3929	Authorized Officer **BARRY SPENCER** Telephone No. (06) 283 2284

INTERNATIONAL PRELIMINARY EXAMIN ON REPORT

International application No.
PCT/AU 96/00149

I. Basis of the report

1. This report has been drawn on the basis of *(Replacement sheets which have been furnished to the receiving Office in response to an invitation under Article 14 are referred to in this report as "originally filed" and are not annexed to the report since they do not contain amendments.)*:

[X] the international application as originally filed.

[ ] the description, pages , as originally filed,
pages , filed with the demand,
pages , filed with the letter of ,
pages , filed with the letter of .

[ ] the claims, Nos. , as originally filed,
Nos. , as amended under Article 19,
Nos. , filed with the demand,
Nos. , filed with the letter of ,
Nos. , filed with the letter of .

[ ] the drawings, sheets/fig , as originally filed,
sheets/fig , filed with the demand,
sheets/fig , filed with the letter of ,
sheets/fig , filed with the letter of .

2. The amendments have resulted in the cancellation of:

[ ] the description, pages

[ ] the claims, Nos.

[ ] the drawings, sheets/fig

3. [ ] This report has been established as if (some of) the amendments had not been made, since they have been considered to go beyond the disclosure as filed, as indicated in the Supplemental Box (Rule 70.2(c)).

4. Additional observations, if necessary:

IV. Lack of unity of invention

1. In response to the invitation to restrict or pay additional fees the applicant has:

- [ ] restricted the claims.
- [ ] paid additional fees.
- [ ] paid additional fees under protest.
- [ ] neither restricted nor paid additional fees.

2. [X] This Authority found that the requirement of unity of invention is not complied with and chose, according to Rule 68.1, not to invite the applicant to restrict or pay additional fees.

3. This Authority considers that the requirement of unity of invention in accordance with Rules 13.1, 13.2 and 13.3 is

- [ ] complied with.
- [X] not complied with for the following reasons:

Claims 41 and 42 are not so limited to an amino acid sequence of a Mycoplasma as to constitute the same invention as claimed in claims 1-40 and 43.

4. Consequently, the following parts of the international application were the subject of international preliminary examination in establishing this report:

- [X] all parts.
- [ ] the parts relating to claims Nos.

V. Reasoned statement under Article 35(2) with regard to novelty, inventive step or industrial applicability; citations and explanations supporting such statement

1. Statement

Novelty (N)	Claims	**1-43**	**YES**
	Claims		**NO**
Inventive step (IS)	Claims	**7-11, 18, 19, 25-32, 34, 40**	**YES**
	Claims	**1-6, 12-17, 20-24, 33, 35-39, 41-43**	**NO**
Industrial applicability (IA)	Claims	**1-43**	**YES**
	Claims		**NO**

2. Citations and explanations

D1 AU,A, 70685/87

D2 AU,B, 49035/90 (640364)

D3 AU,A, 76820/91

D4 AU,A, 17602/95

D5 AU,B, 49599/90 (638970)

D6 US,A, 4894332

D7 US,A, 5252328

D8 US,A, 5240706

D9 EP,A, 0475185

D10 Journal of Bacteriology, Volume 177, No. 7, pages 1915-1917

D11 Journal of Clinical Microbiology, Volume 33, No. 3, pages 680-683

D12 Infection and Immunity, Volume 49, No. 2, pages 329-335

Claims 1-6, 12-17, 20-24, 33, 35-39, 41-43 do not involve an inventive step when compared to D2, when read in the light of D1, D3-D12. D1, D3-D12 show that before the priority date, the use of antigens targeted to surface lipoproteins or membrane proteins of Mycoplasma were known and sought after. D2 discloses the exact same method for a group of pathogens, one of which can be mycoplasmas. A PSA having regard to all these documents would find no burden of experimentation or inventive faculty to use the same method as D2 for obtaining antigens to mycoplasmas.

D1 discloses taking a biological sample three days after the last injection which would be included within your term "short time".

D10-D12 all disclose the nucleic acid sequences as set out in claims 14, 15, 38 and 42.

VI. Certain documents cited

1. Certain published documents (Rule 70.10)

Application No. Patent No.	Publication date *(day/month/year)*	Filing date *(day/month/year)*	Priority date (valid claim) *(day/month/year)*
AU,A, 17602/95	26 October 1995	21 April 1995	2 April 1990

2. Non-written disclosures (Rule 70.9)

Kind of non-written disclosure	Date of non-written disclosure *(day/month/year)*	Date of written disclosure referring to non-written disclosure *(day/month/year)*

VIII. Certain bservations on the international application

The following observations on the clarity of the claims, description, and drawings or on the question whether the claims are fully supported by the description, are made:

Claims 1 and 16 lack clarity in their reference to "short time". It is unclear what time frame is included by this term and the description gives no further indication as to any limits.

Claims 41 and 42 are not fully supported by the description because the description is directed towards Mycoplasma yet these claims are to any amino acid sequence.